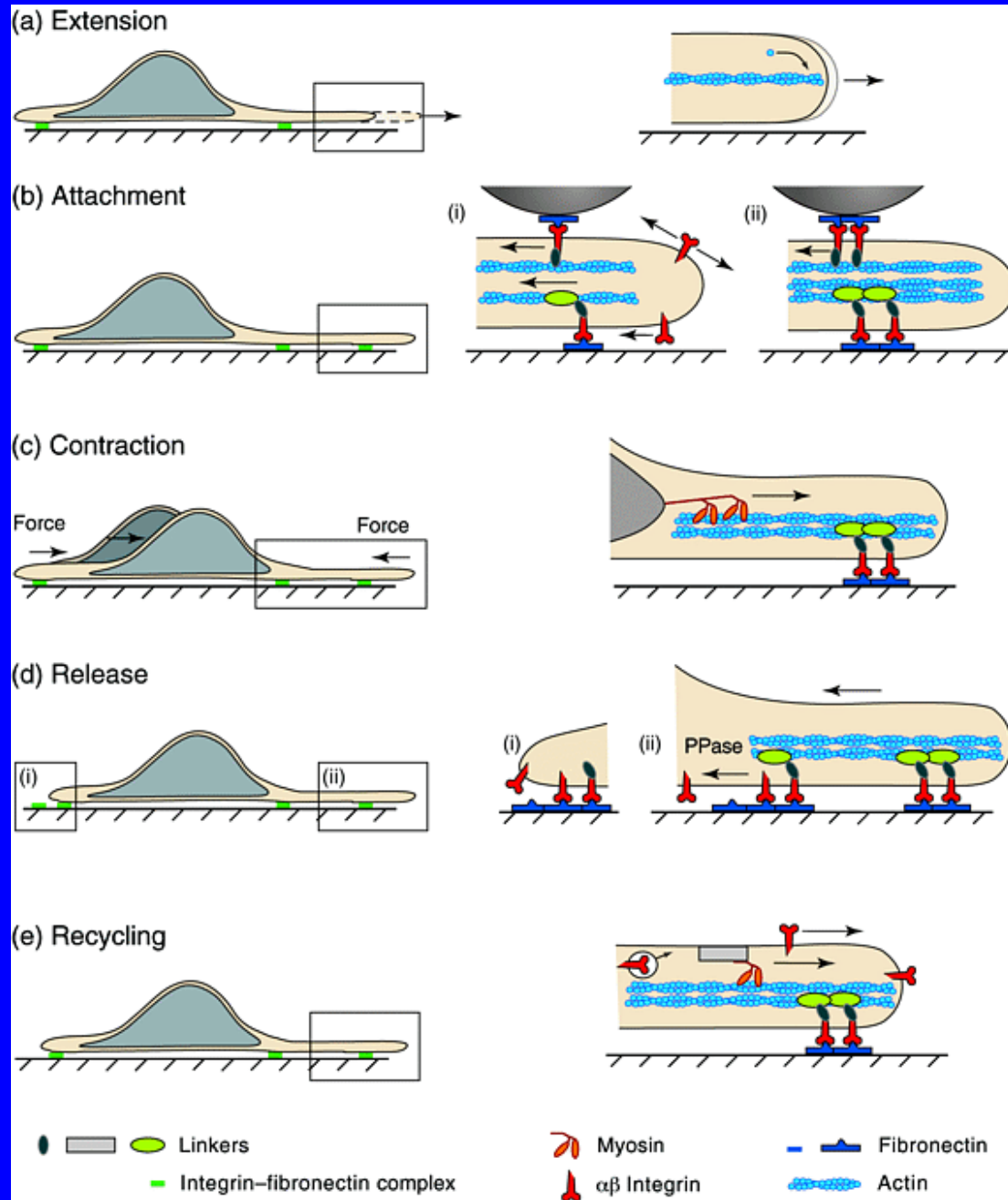


# Mechanical Sensing in Normal and Transformed Cells

- What differentiates a formed organism from a sphere of cells is the ability to apply and sense forces at specific places and times
- Rapid neuronal sensing of force is through ion channels. Turgor in tissues and long-term forces are sensed by cytoskeletal-based mechanisms.
- Transformation is described as the ability to grow on soft agar, i.e. in the absence of force.

# Cell migration: a cyclic process of force generation

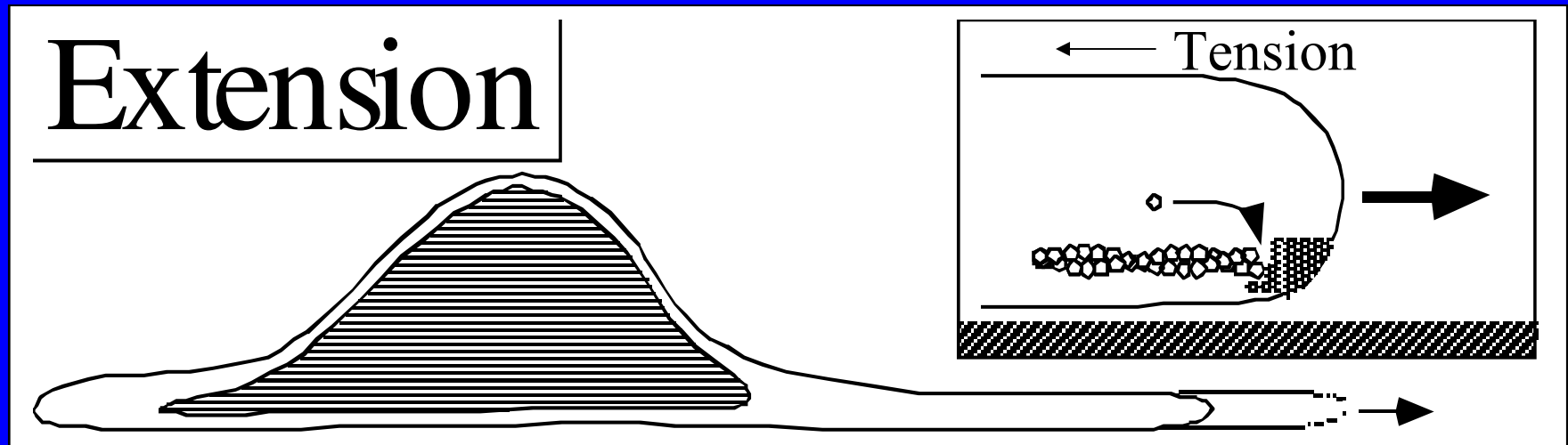


Membrane-Cyto. Adhesion Resists Extension  $\alpha$  PIP2. Sheetz, Nature Rev. MCB. 2:392 (2001)

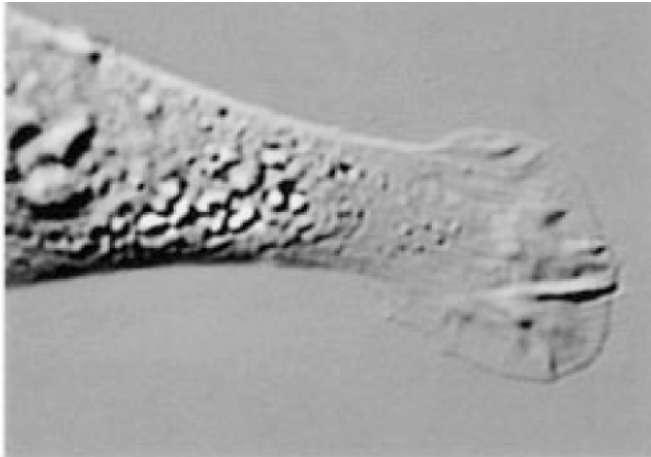
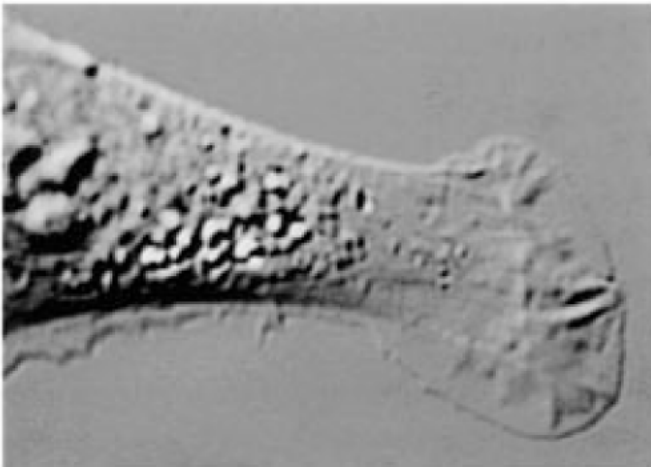
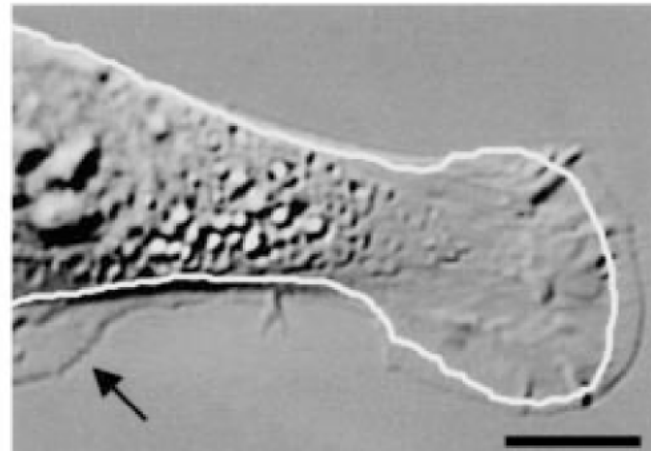
Extracellular Matrix Adhesion Linked to Force Generation by Rearward Flow

How do cells locally transduce Matrix rigidity into a signal used to direct cell edge extension?

Apparent Membrane Tension  
Resists Extension by Actin and  
Tension  $\alpha$  [PIP2]



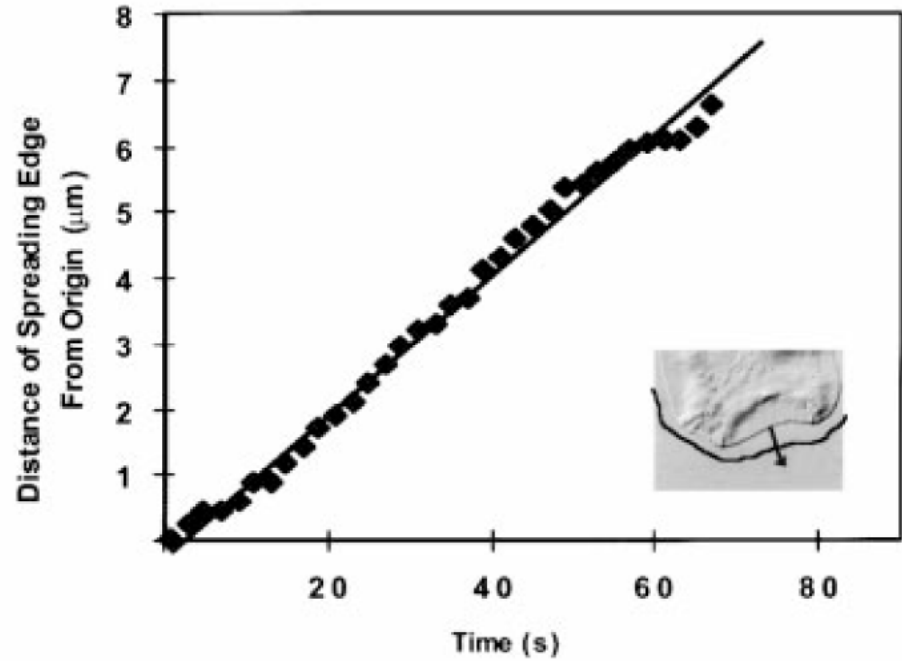
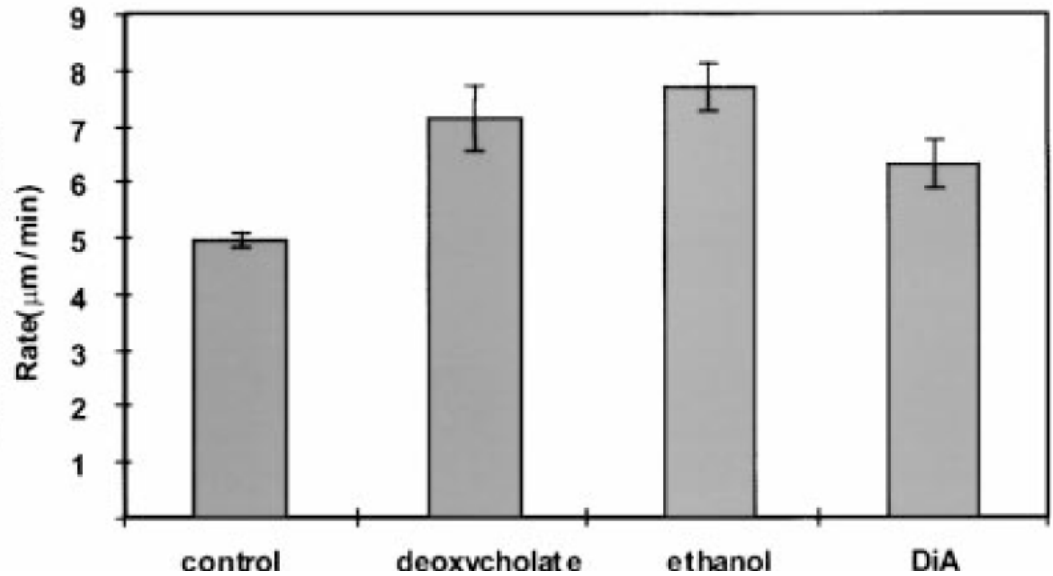
Reviewed: Sheetz, Nature Rev. MCB. 2:392 (2001)

**a****b****c**

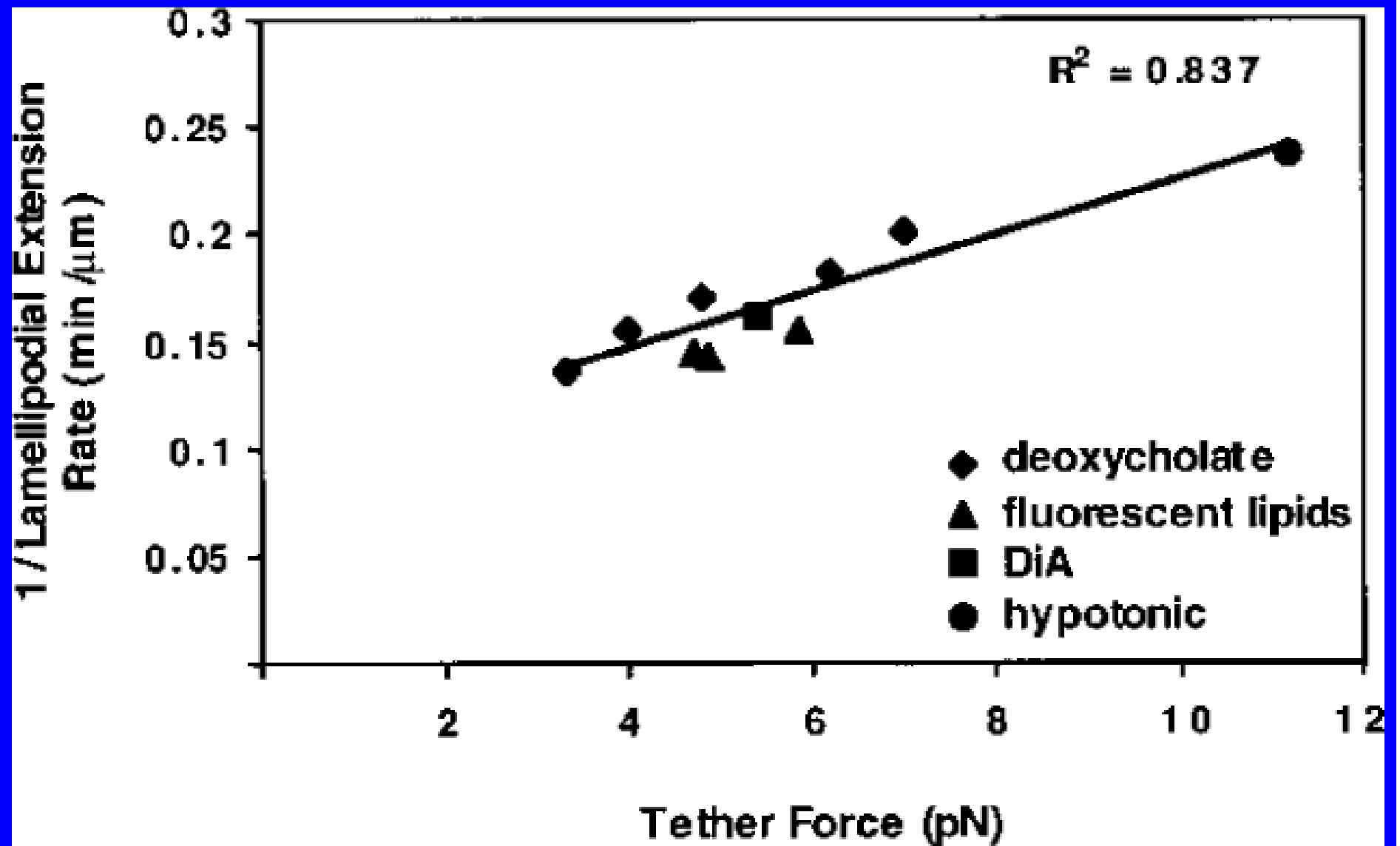
counted, normal number of lamellipodia

*Reagents that Increase Membrane Area Increase Lamellipodial Extension*

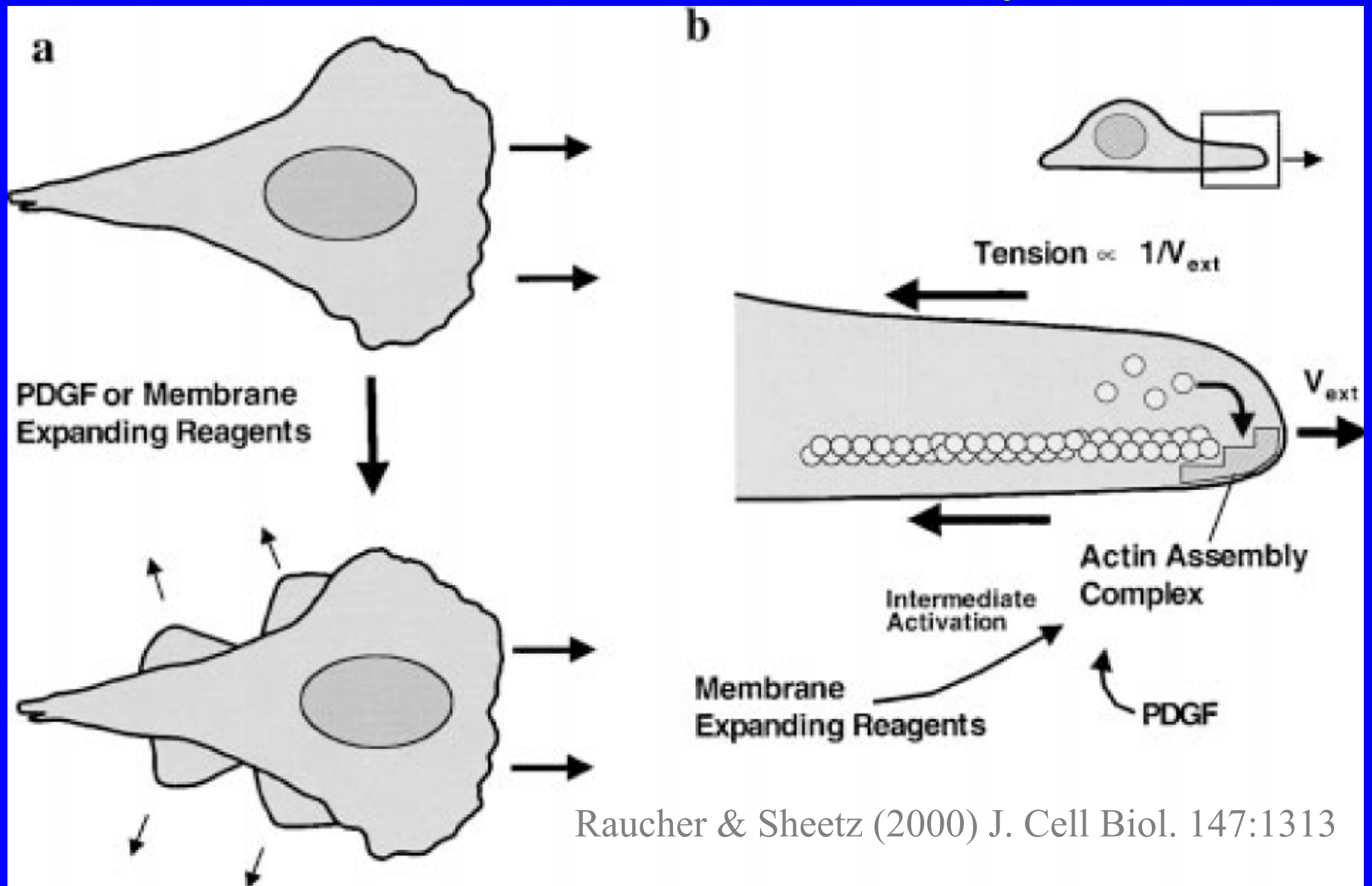
position at the (inset). The recovery over  $t^2 > 0.5$ . (e) Division of the cho-mM bars mea-

**d****e**

Lamellipodial Extension



# Membrane Load Alters Activation and Rate of Actin Assembly

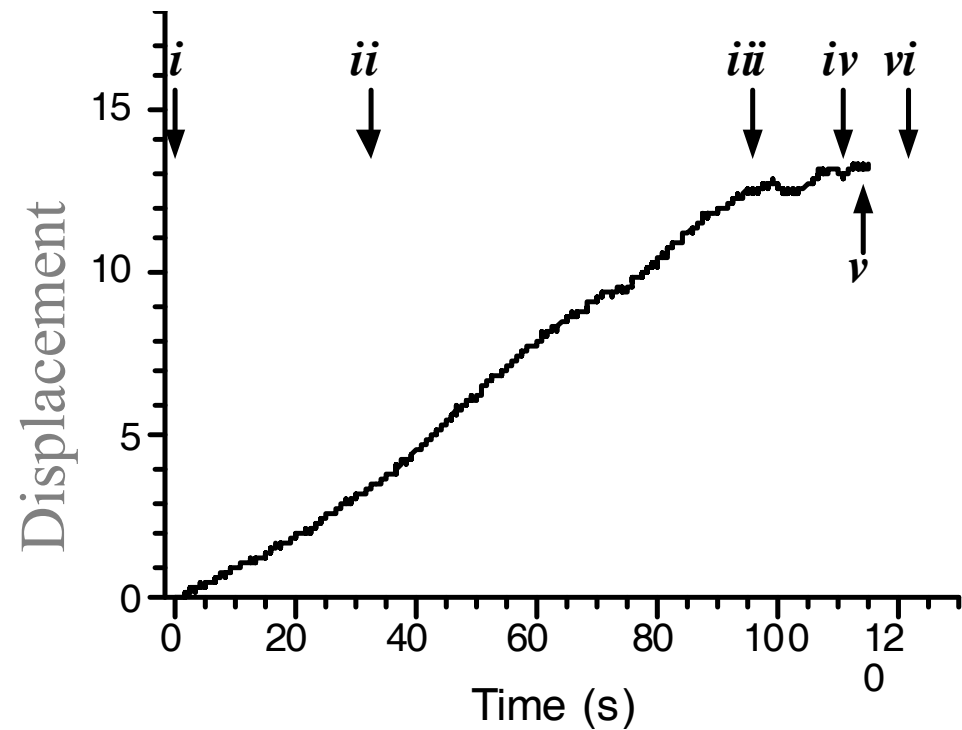
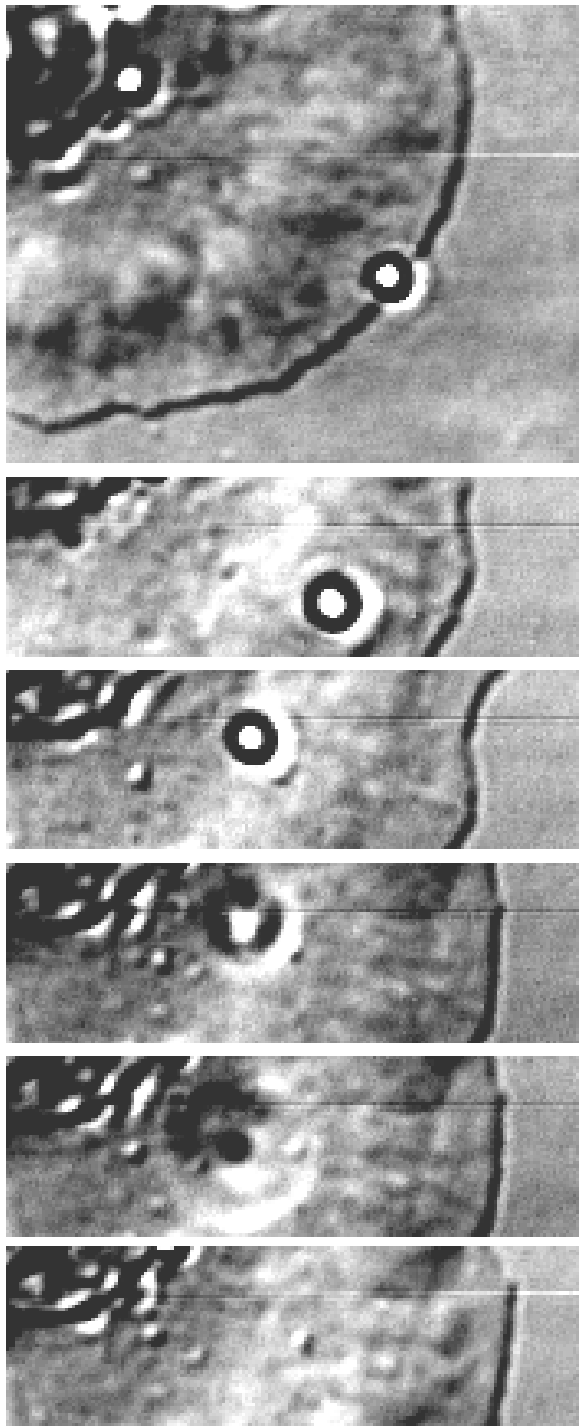


Raucher & Sheetz (2000) J. Cell Biol. 147:1313

# Membrane Tension: Physical Control of Cell Functions

- Adhesion of membrane to the cytoskeleton develops tension that provides general regulation of endocytosis rate (surface-area/volume ratio), motility, and resealing
- Model of lipid-dependent membrane-cytoskeleton adhesion
- PIP2 is major lipid controlling membrane-cytoskeleton adhesion
- Tension is too low to activate stretch channels

# FN Bead Binding Movement and Release

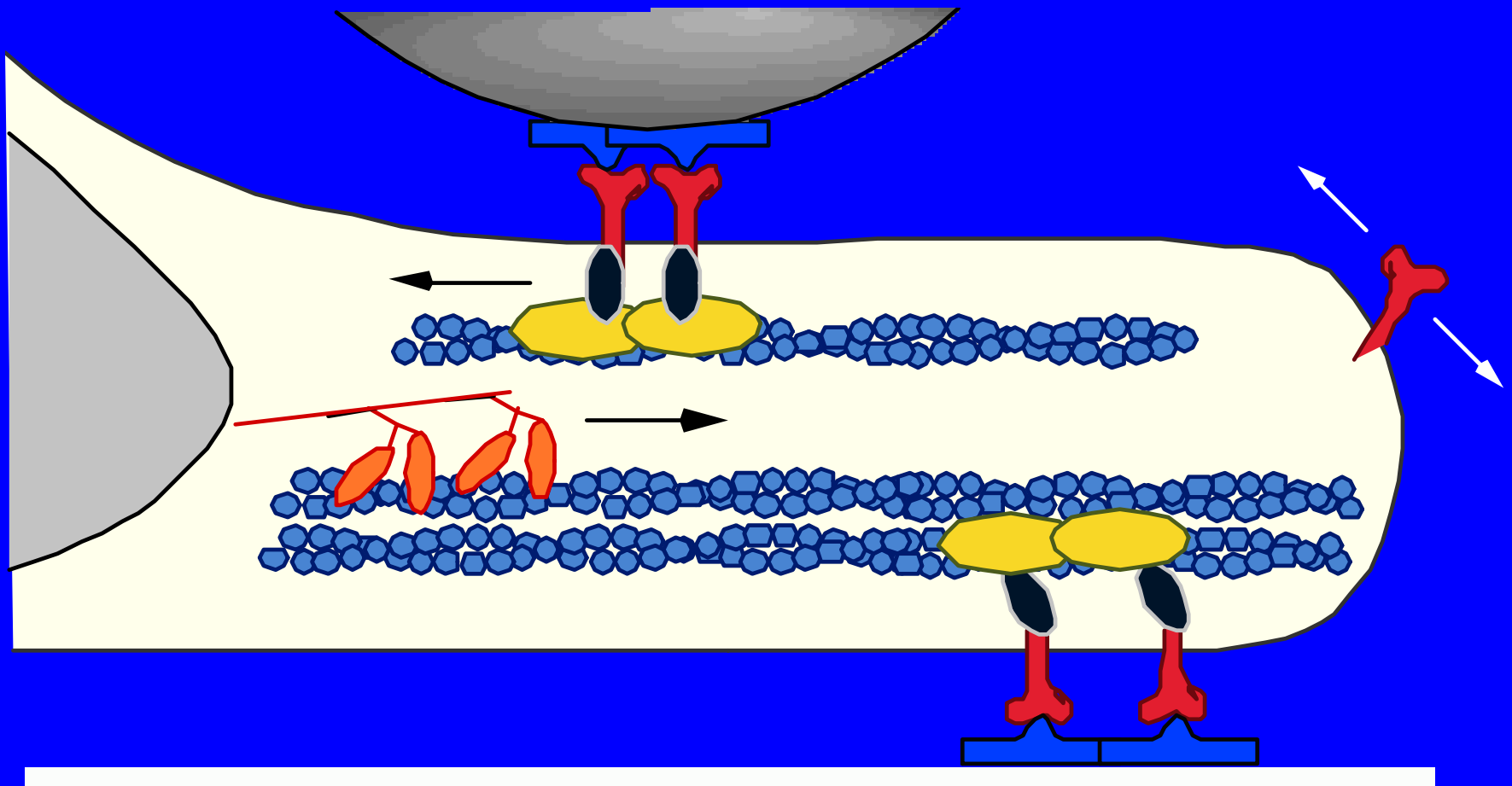


Nishizaka et al., PNAS 97:692 (1999)

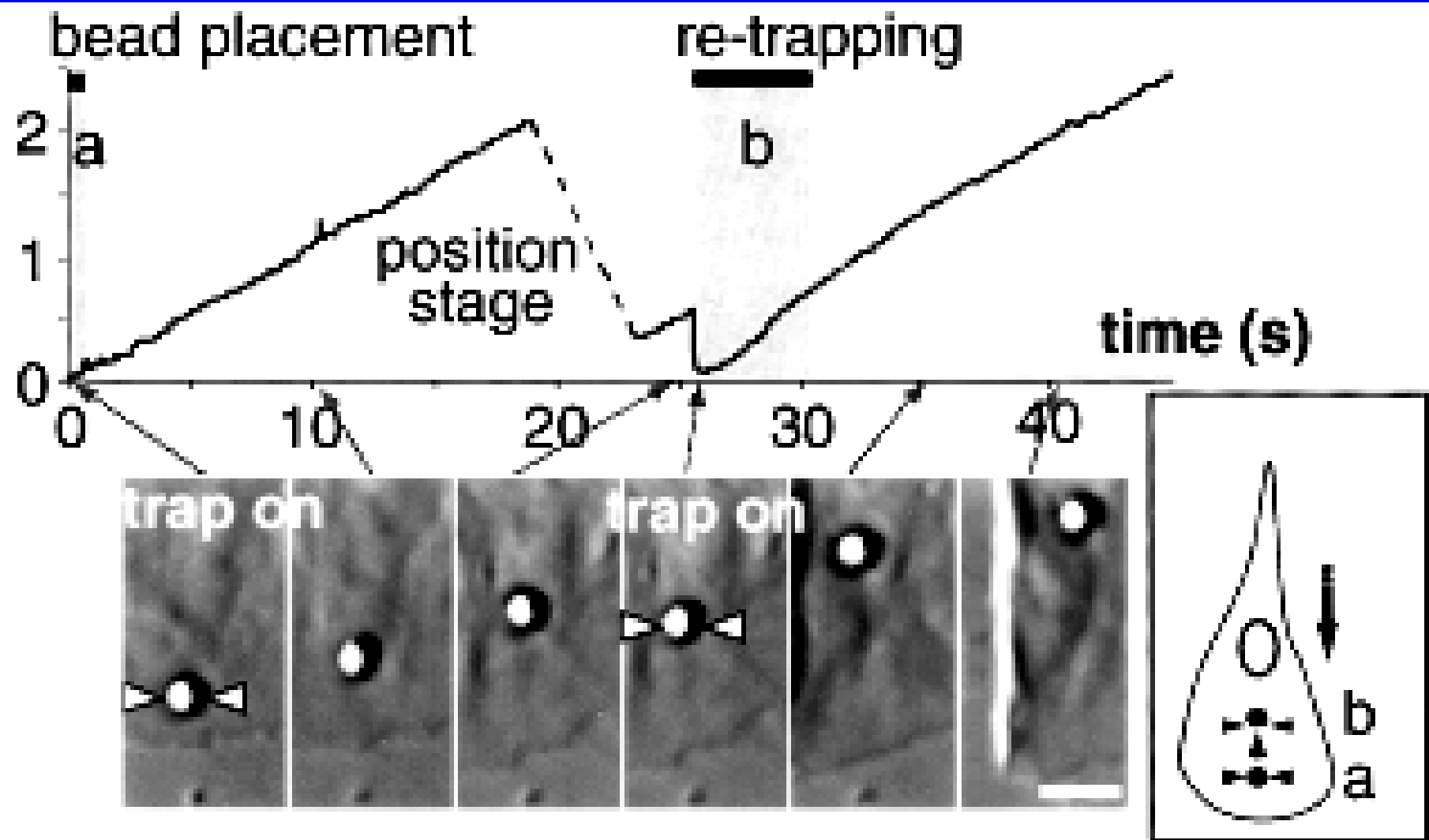


How does a cell treat the ECM?

# Attachment and Contraction

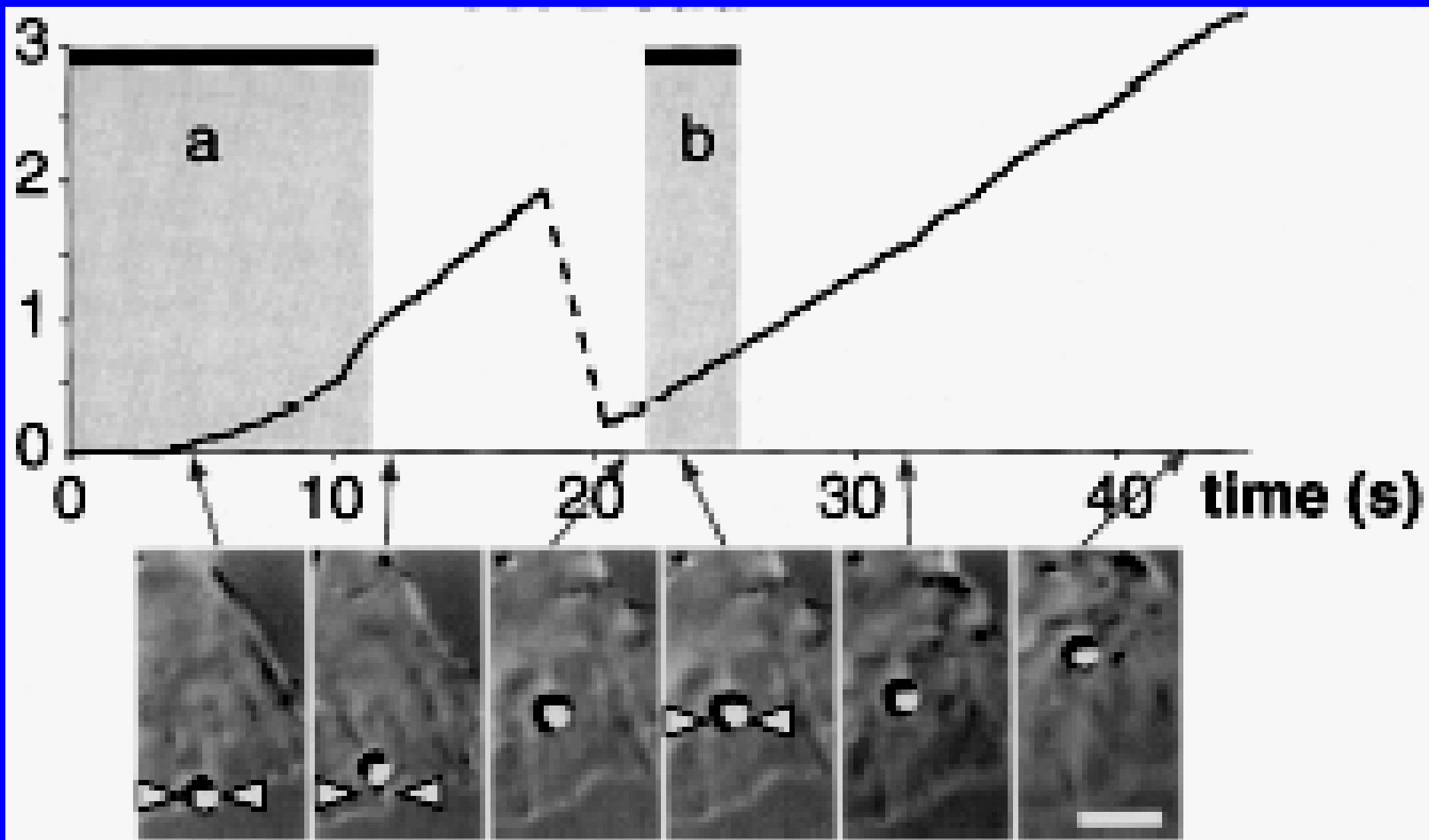


# Unstressed Attachments



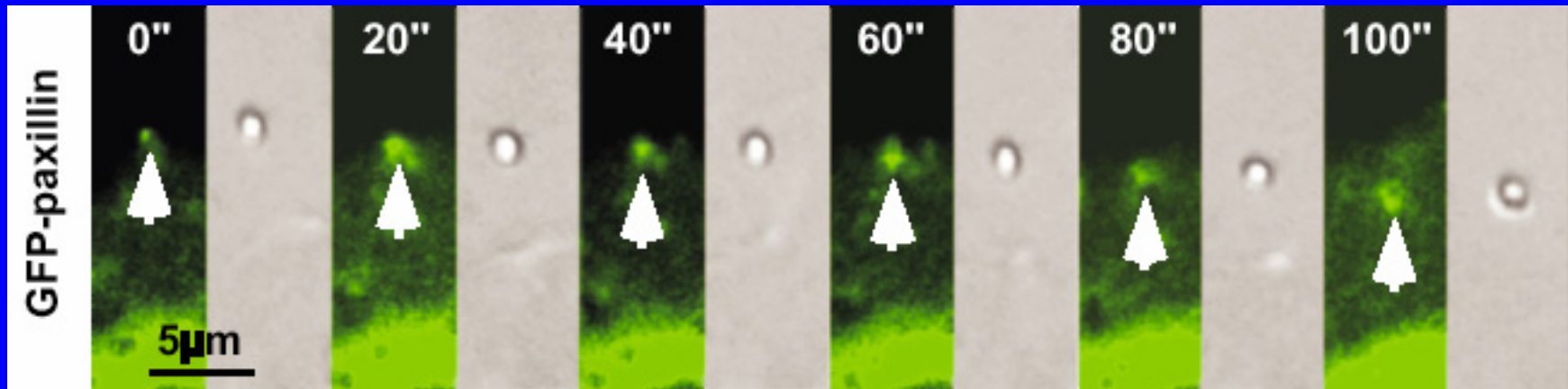
Choquet et al., Cell. 88:39 (1997)

# Force Causes Reinforcement



# Force Causes GFP-Paxillin

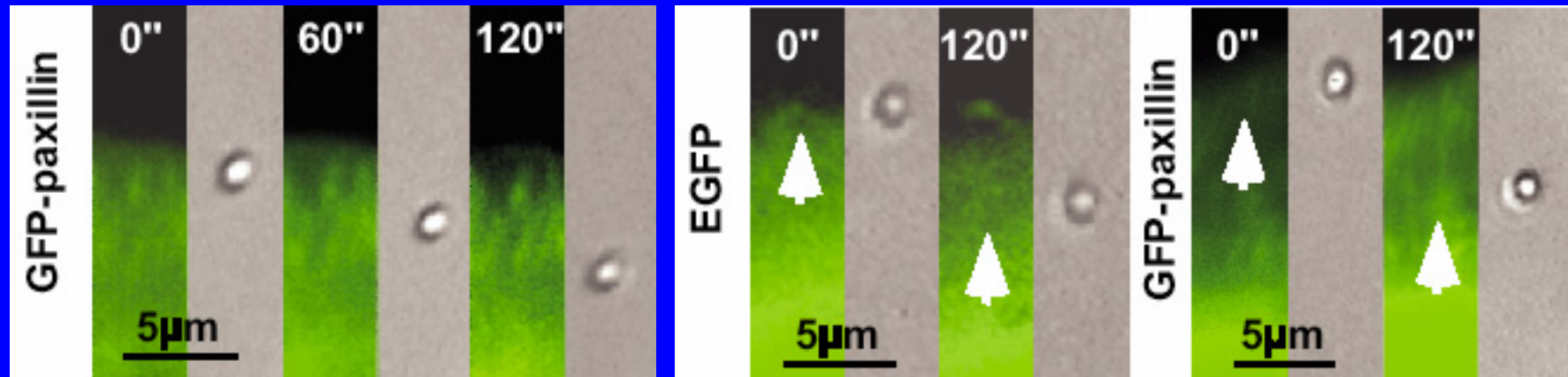
## Accumulation in RPTP $\alpha$ <sup>+/+</sup> with FN



Without Force

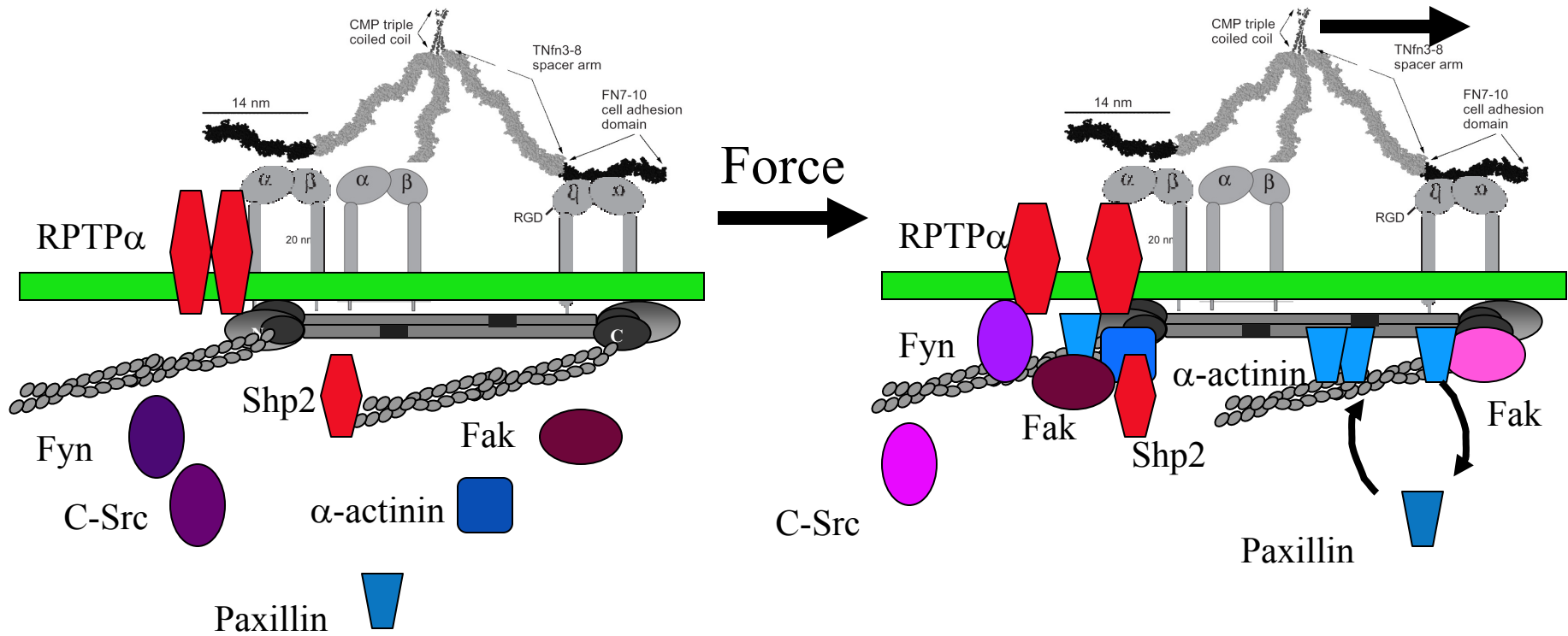
FN Bead

Con A Bead



Von Wichert et al., 2003. J. Cell Biol. 161:143-153

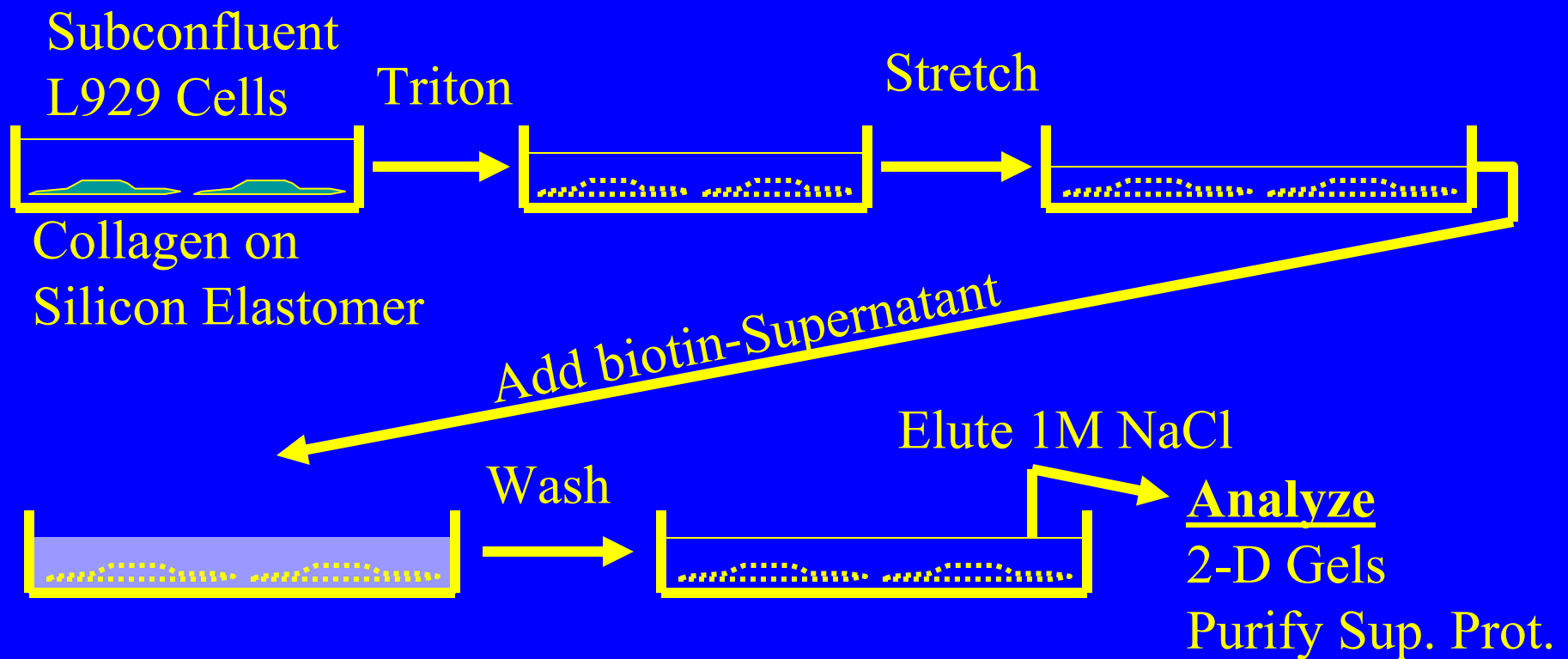
# Force Effects in FN Contacts



# Cytoskeleton Stretching Causes Focal Contact Protein Binding

We tested the hypothesis that the cytoskeleton is being altered directly by force on matrix molecules using a stretchable silicon substrate and detergent-treated cells.

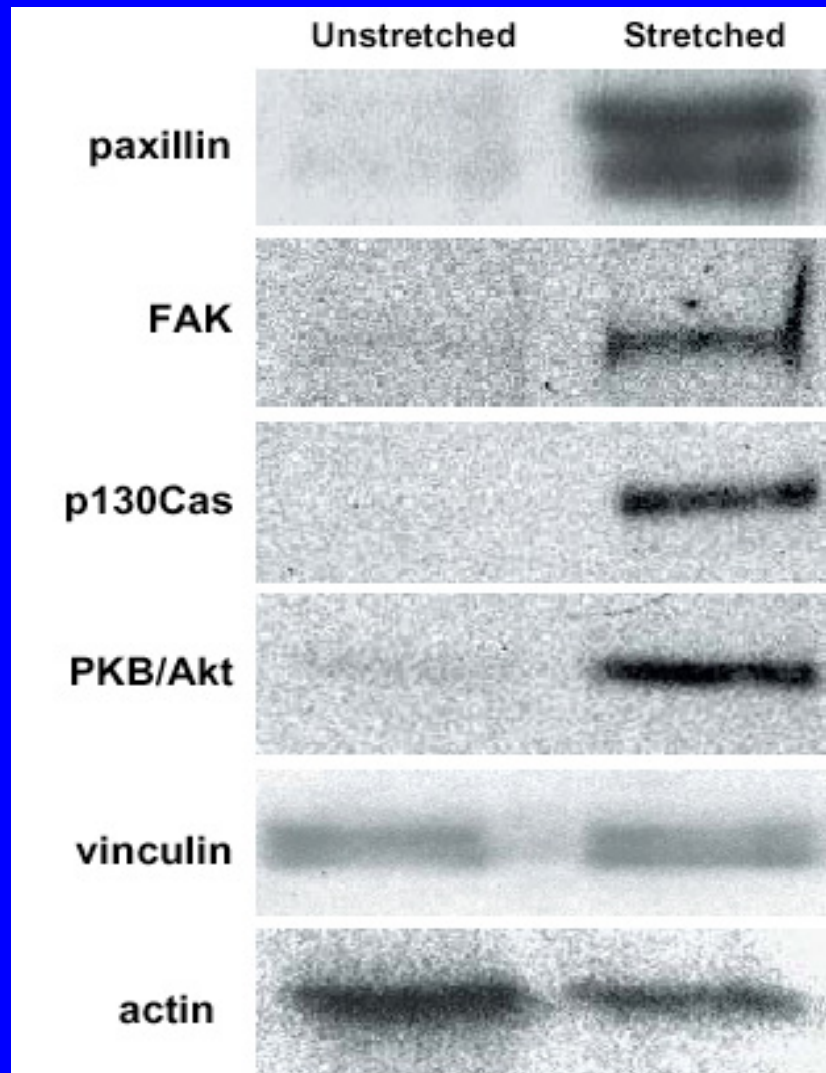
# Stretching of Cytoskeletons



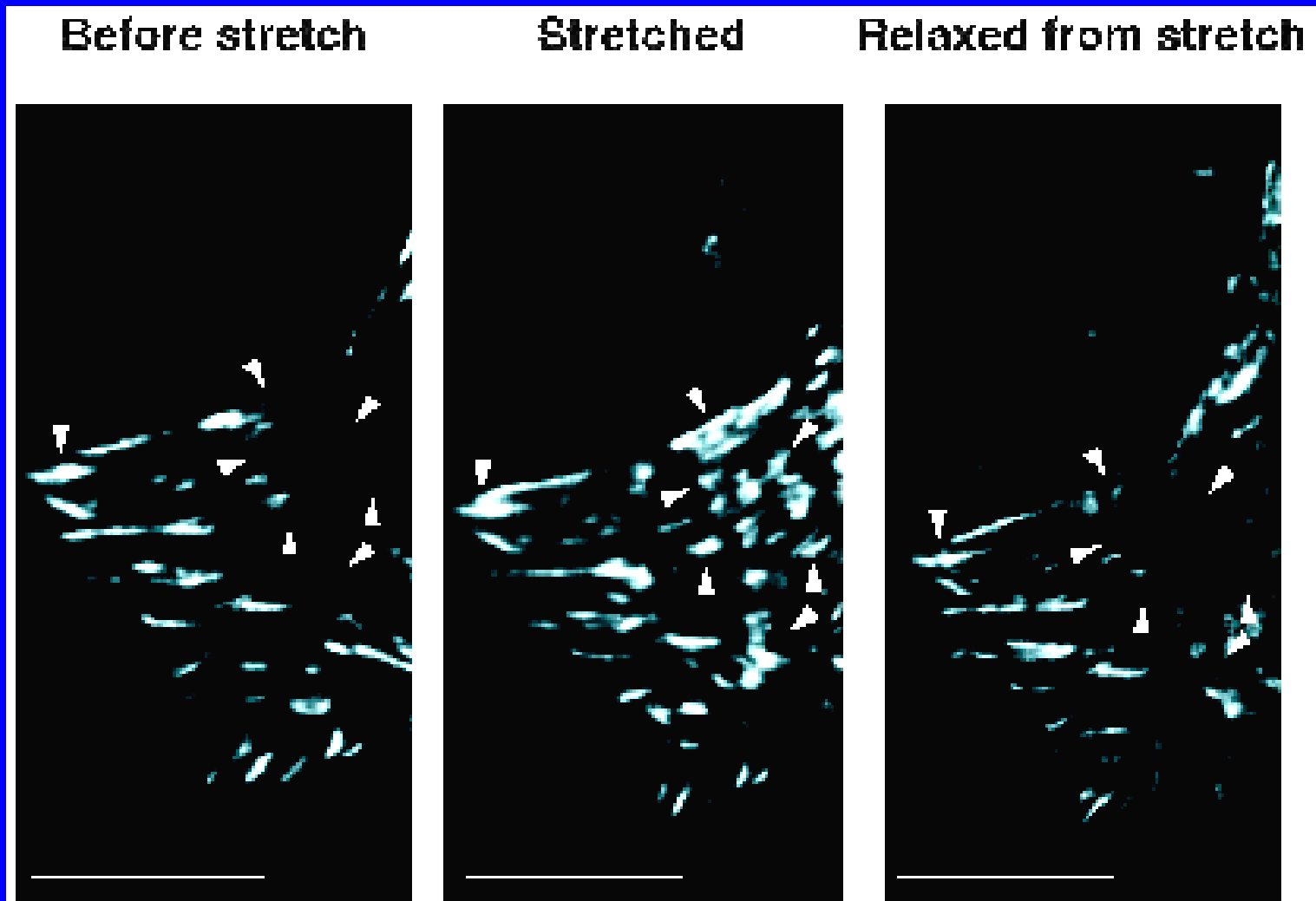
Sawada and Sheetz, JCB **156**:609 (2002)



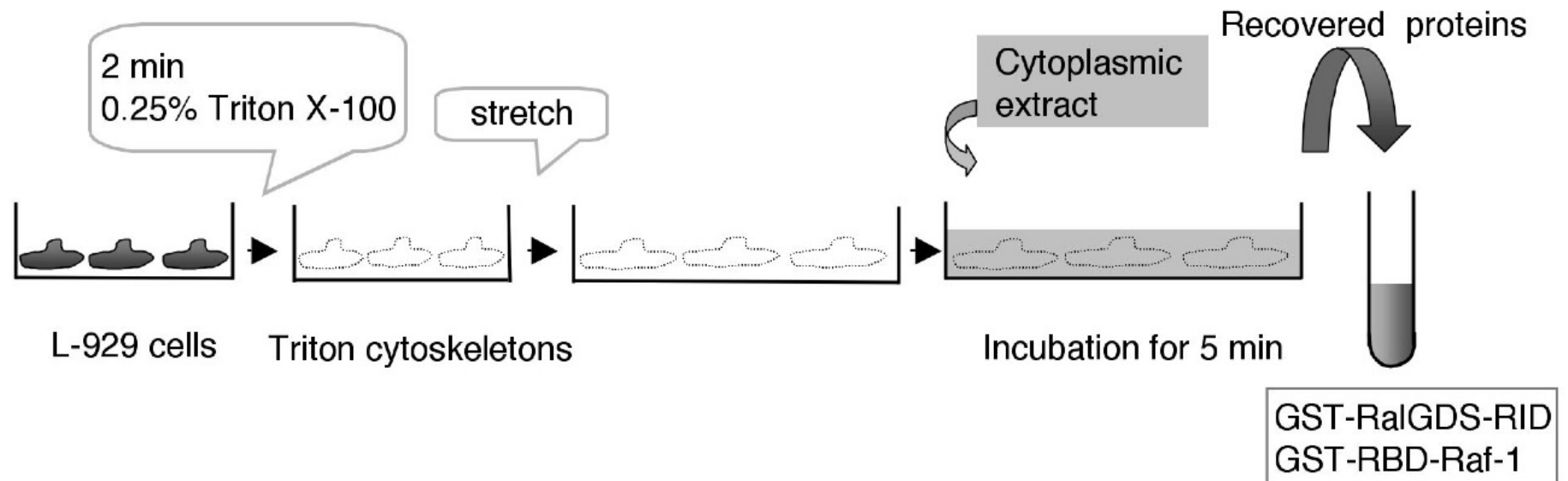
# Antibody Analysis of Bound Proteins



# GFP-Paxillin Stretch-Dep. Binding

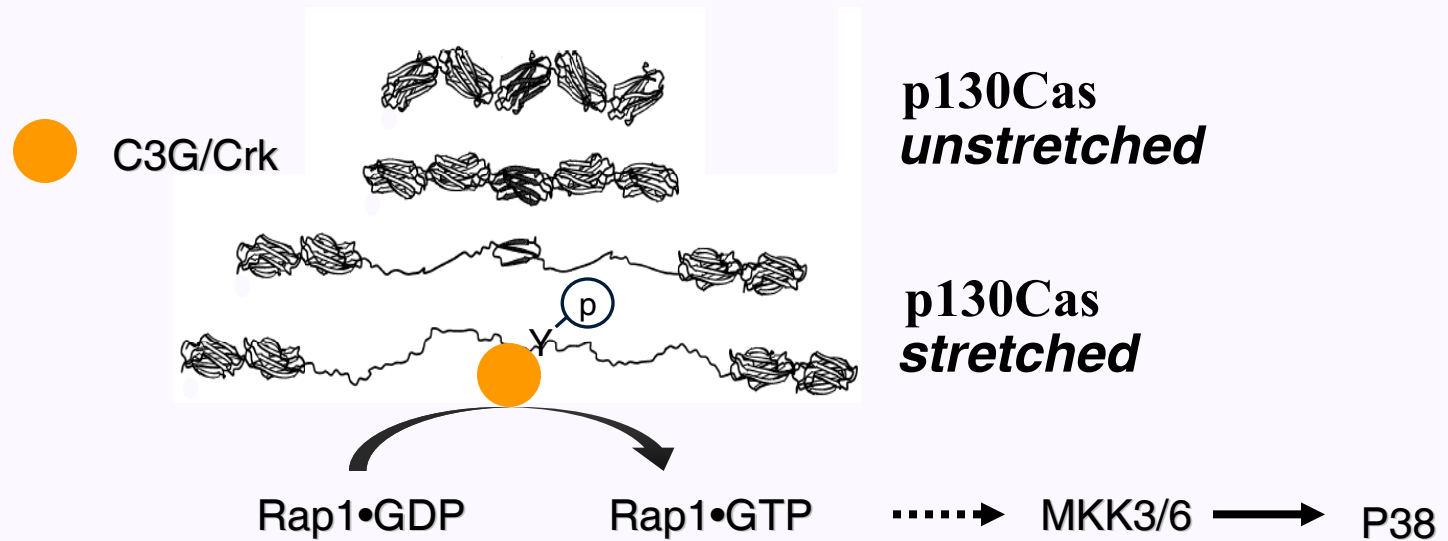


# Protocol for Measuring G Protein Activation by Cytoskeletons

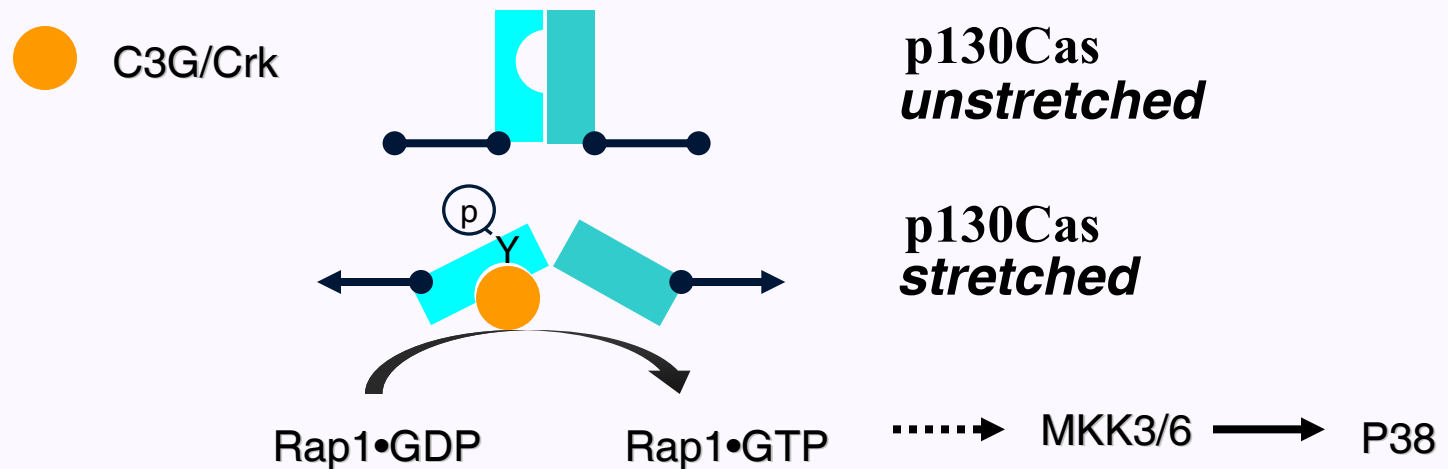


Tamada et al., 2004. *Develop Cell*, 7:709-18

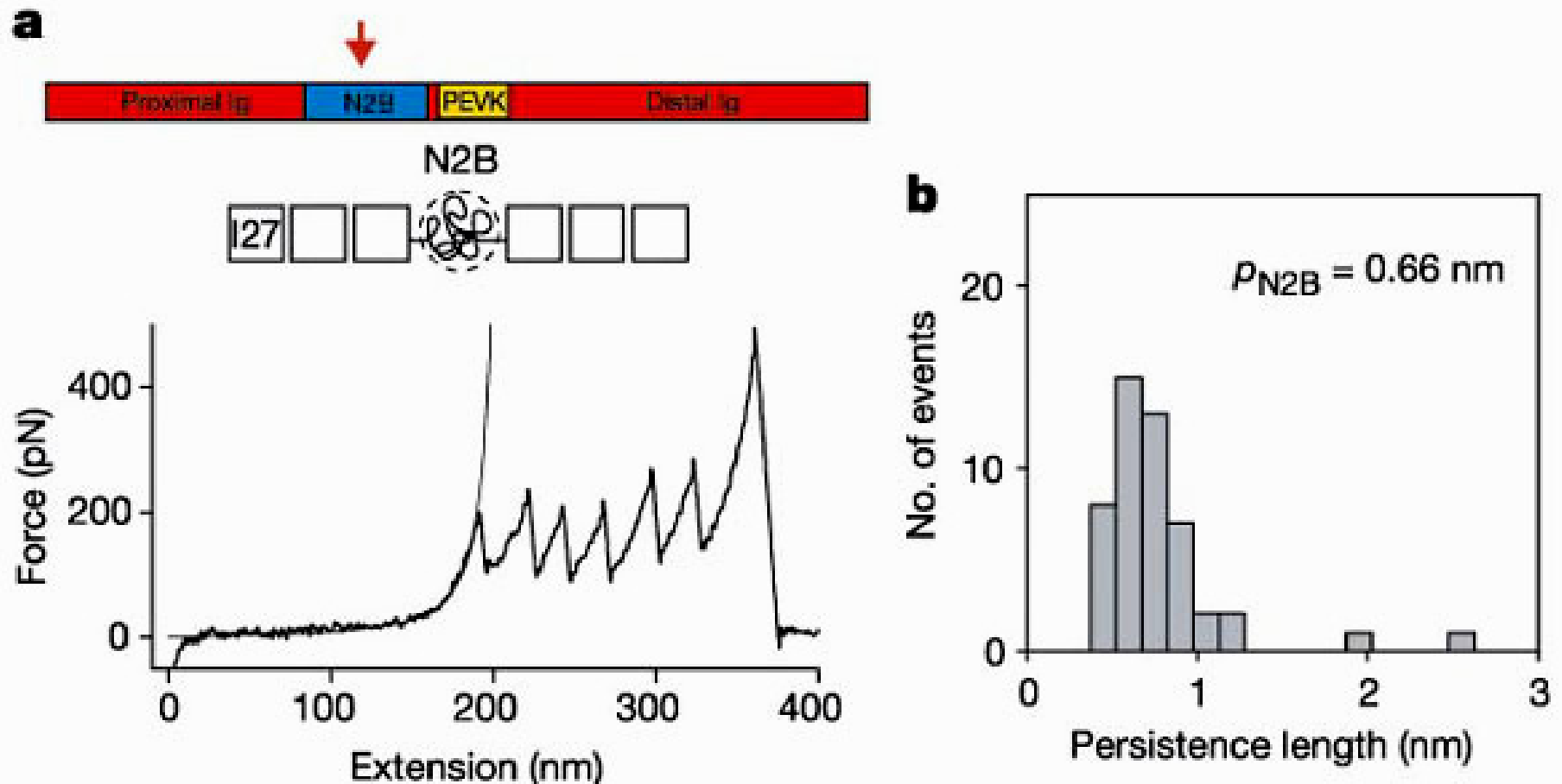
### (A) Protein Unfolding



### (B) Protein Distortion

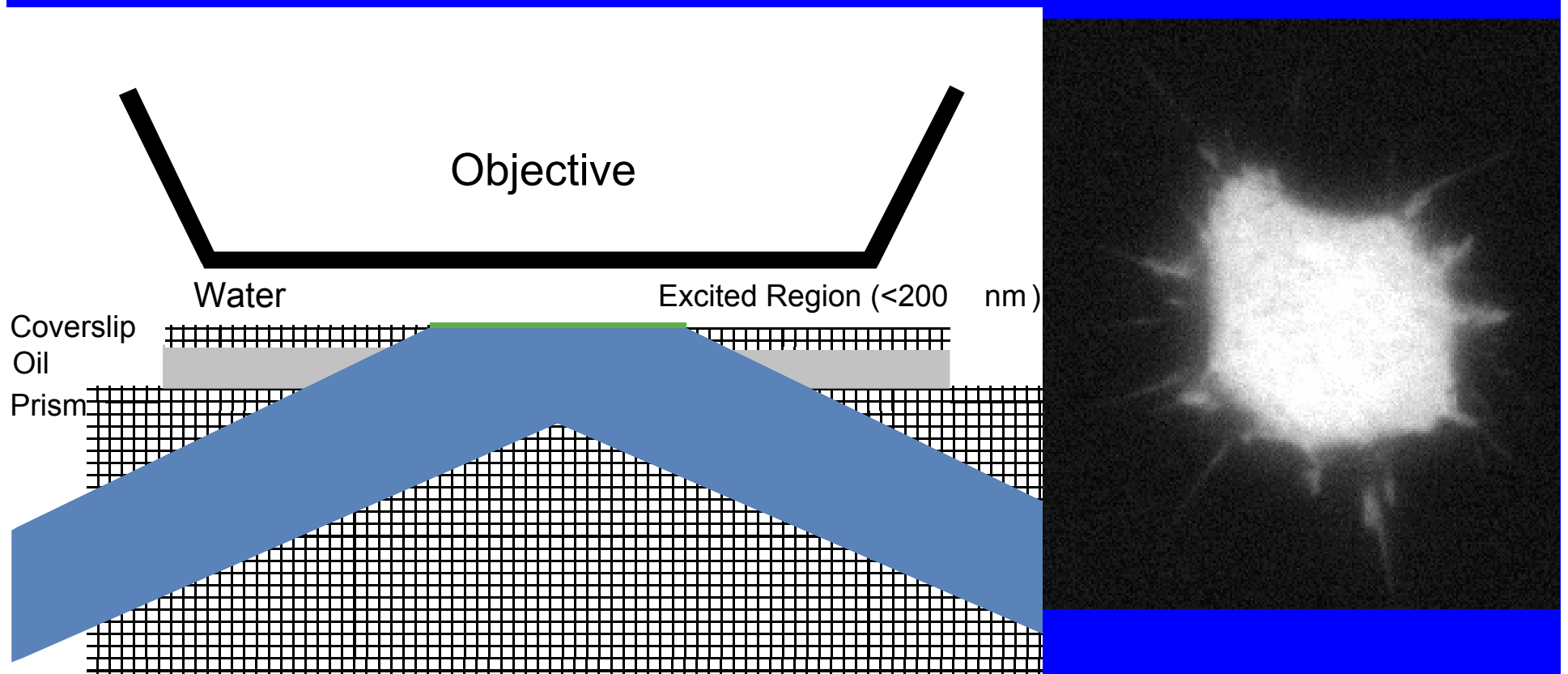


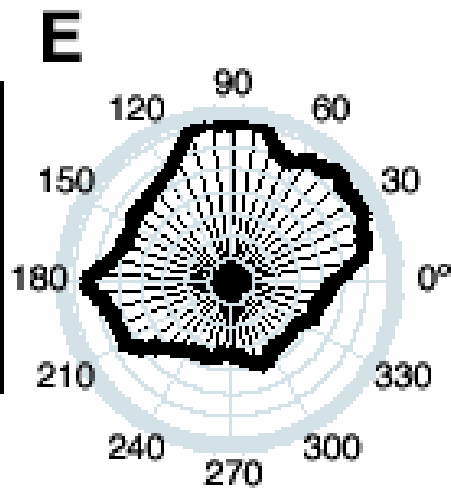
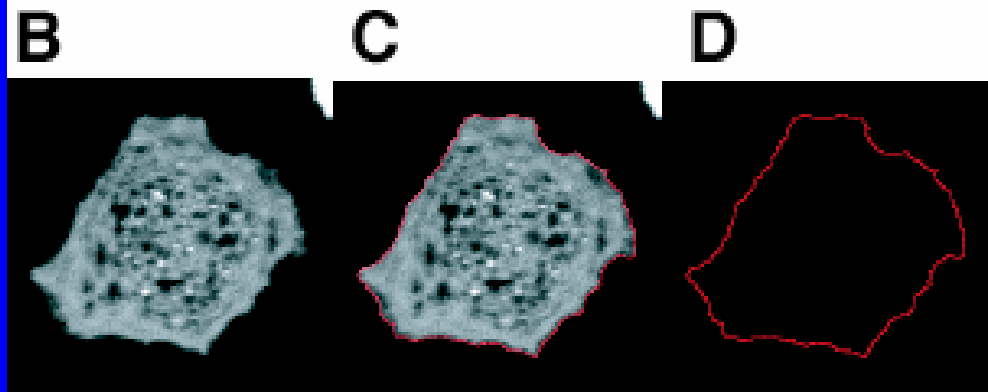
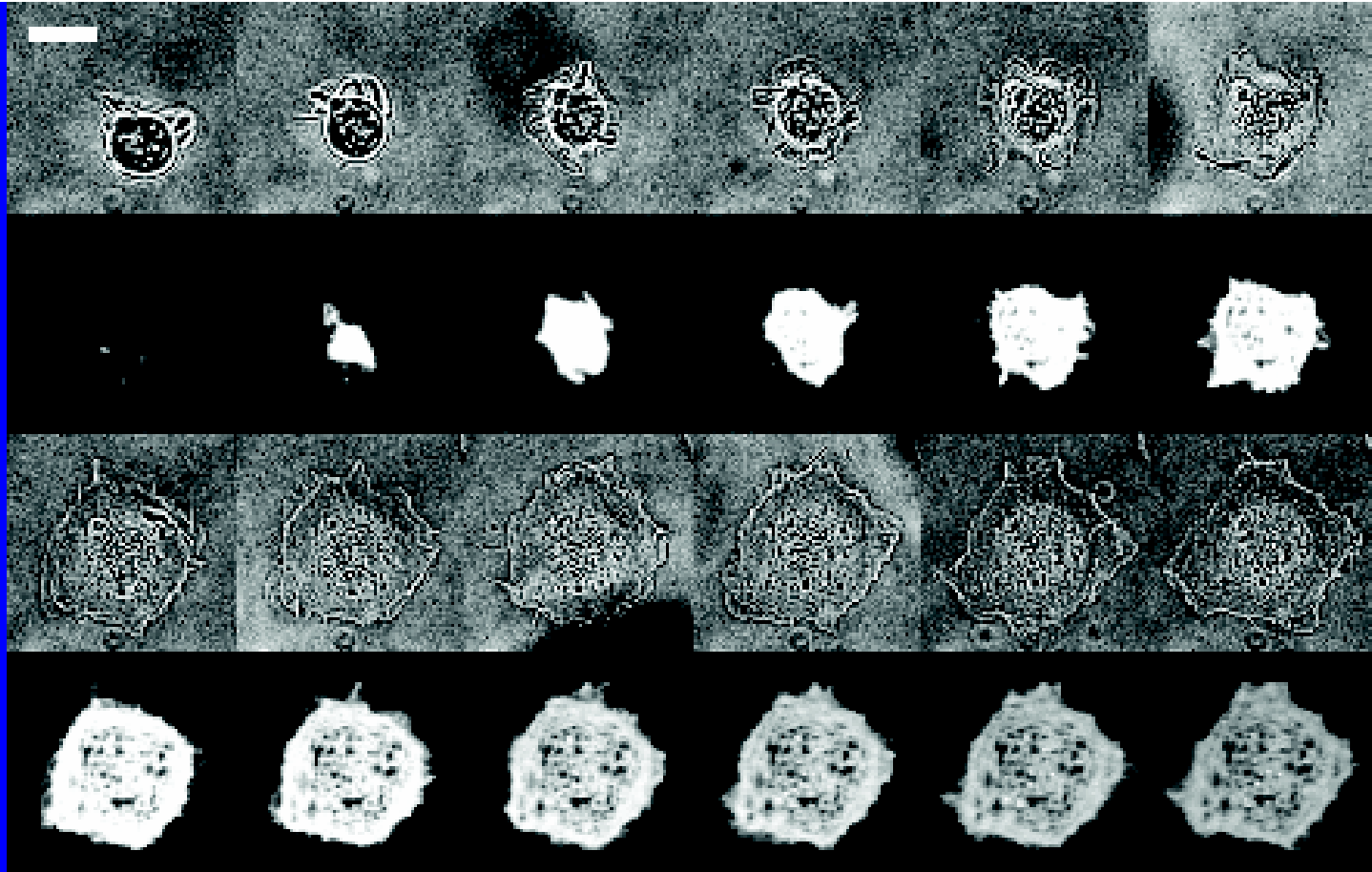
# Titin Domains Unfold as Expected for a Force Sensor

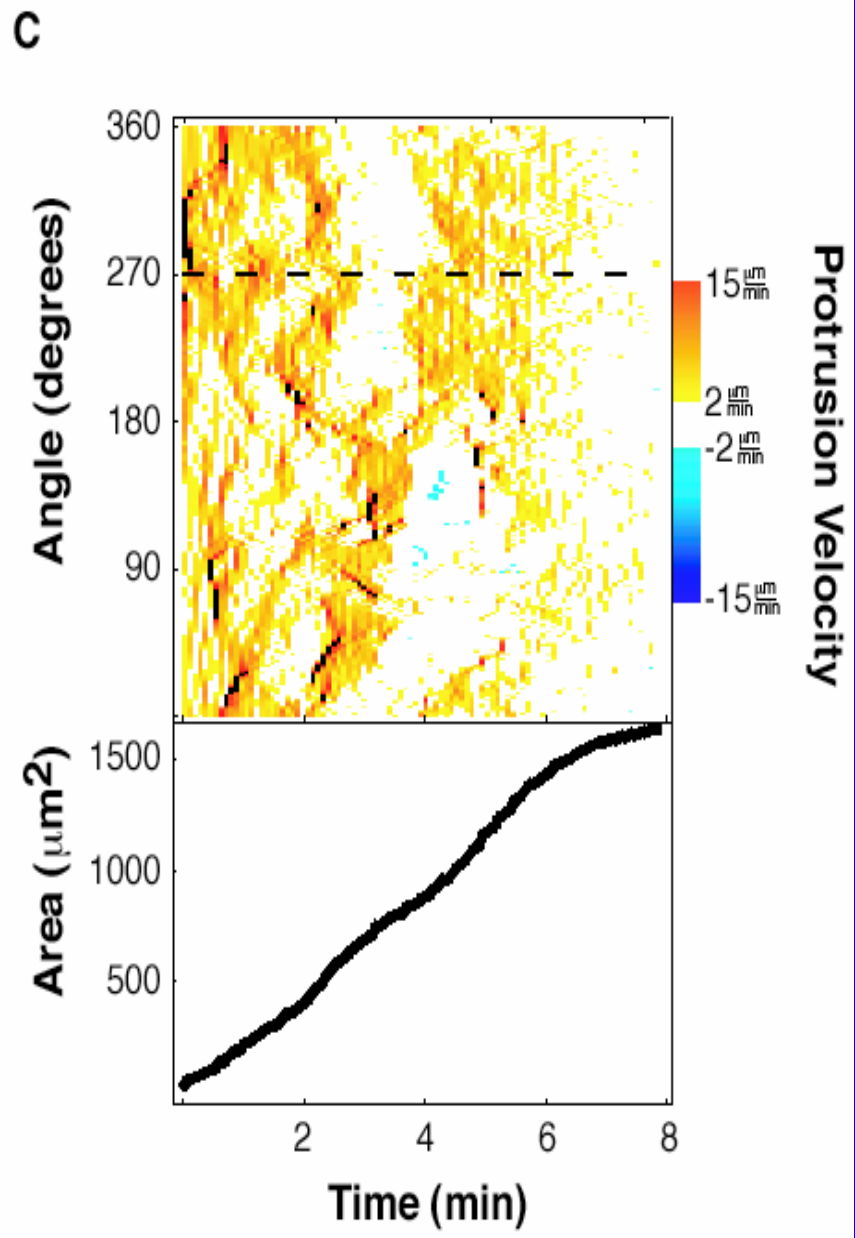
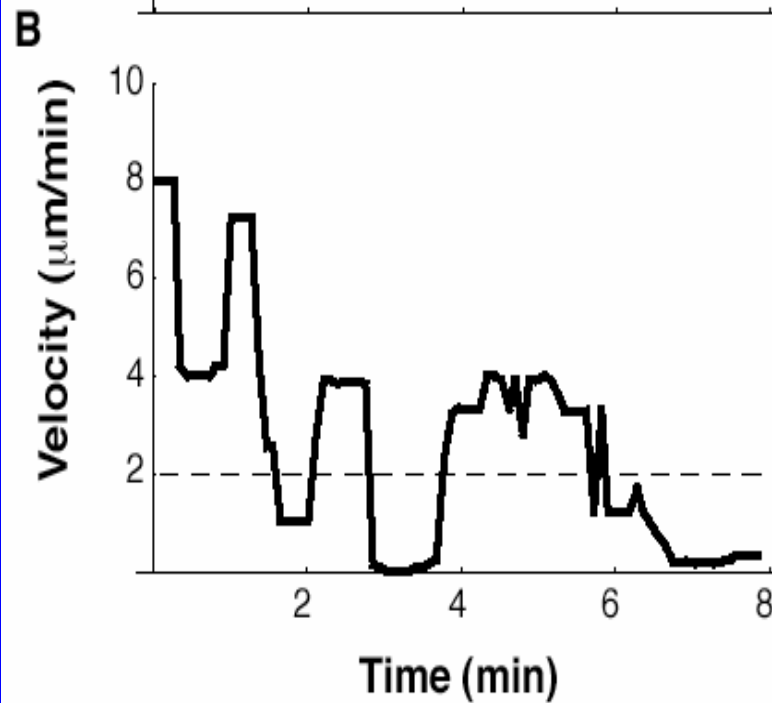
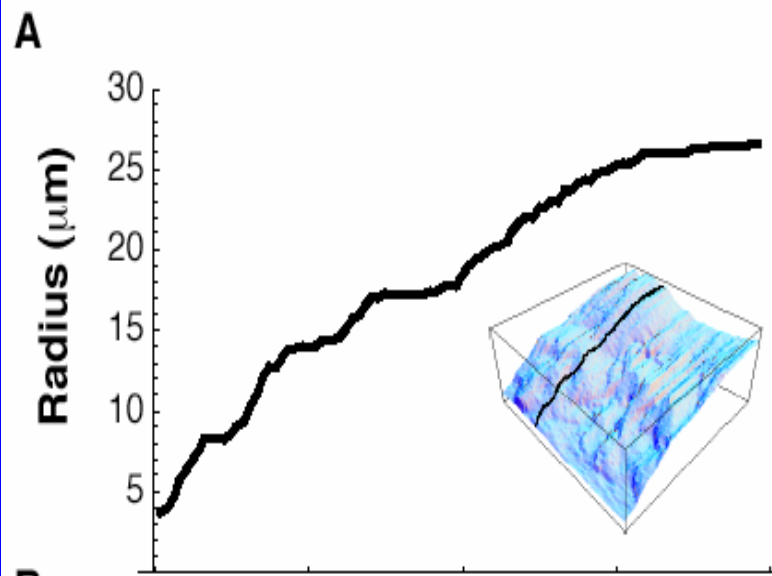


Li et al., (2002) Reverse Engineering of Titin. Nature, 418:998-1002

# Total Internal Reflection Fluorescence of Calcein-loaded Cell Spreading

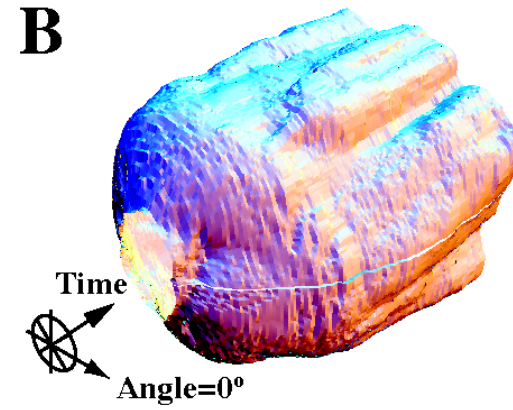
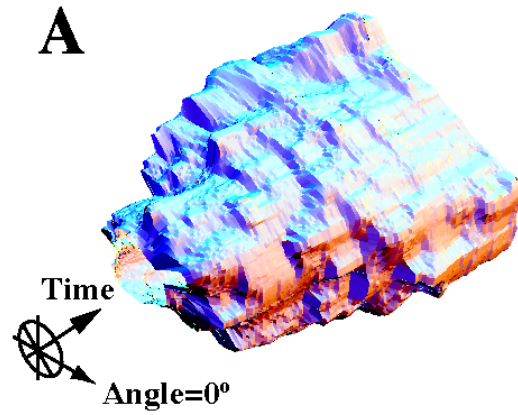






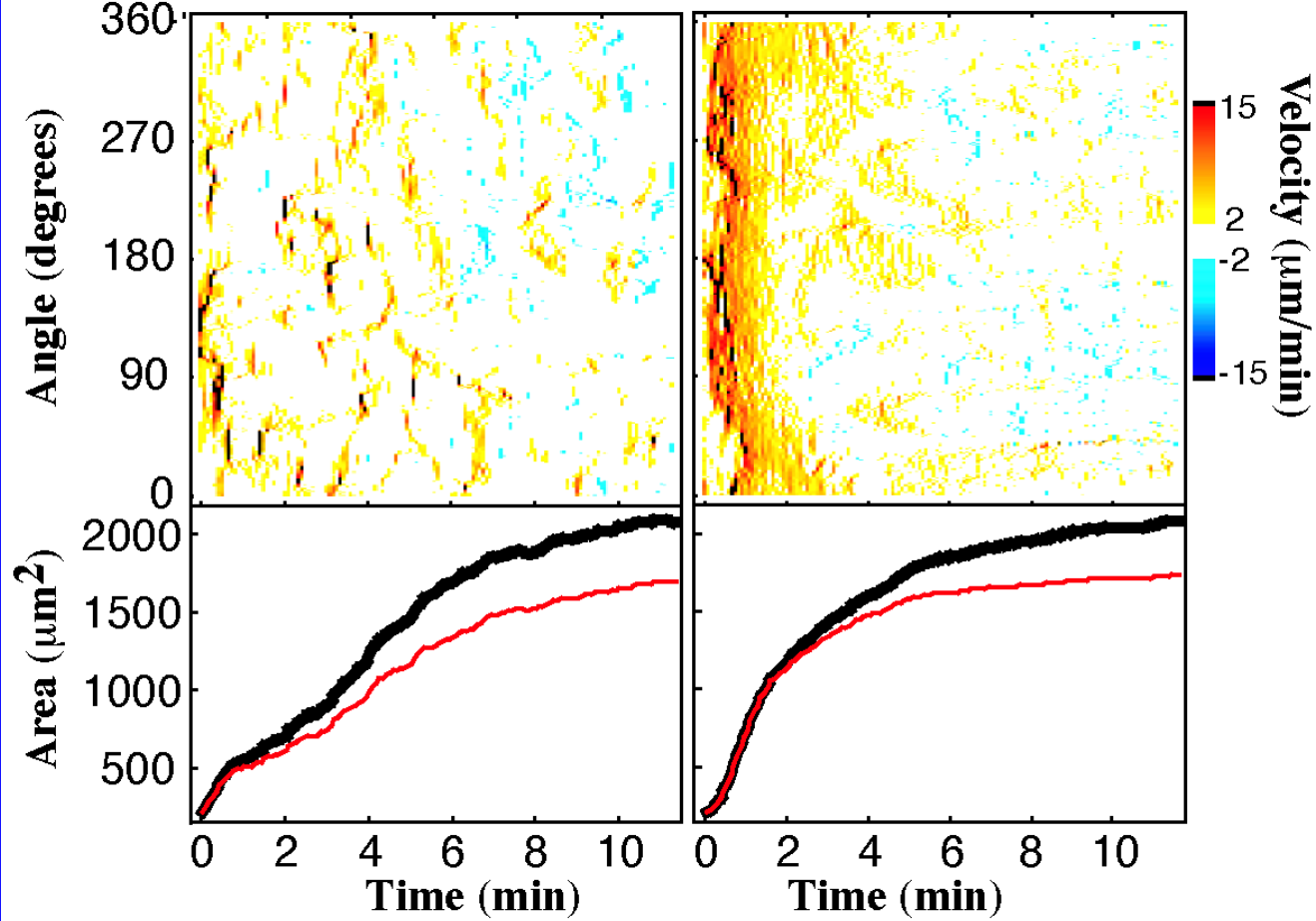
Dubin-Thaler et al., Biophys. J, 86:1794 (2004).



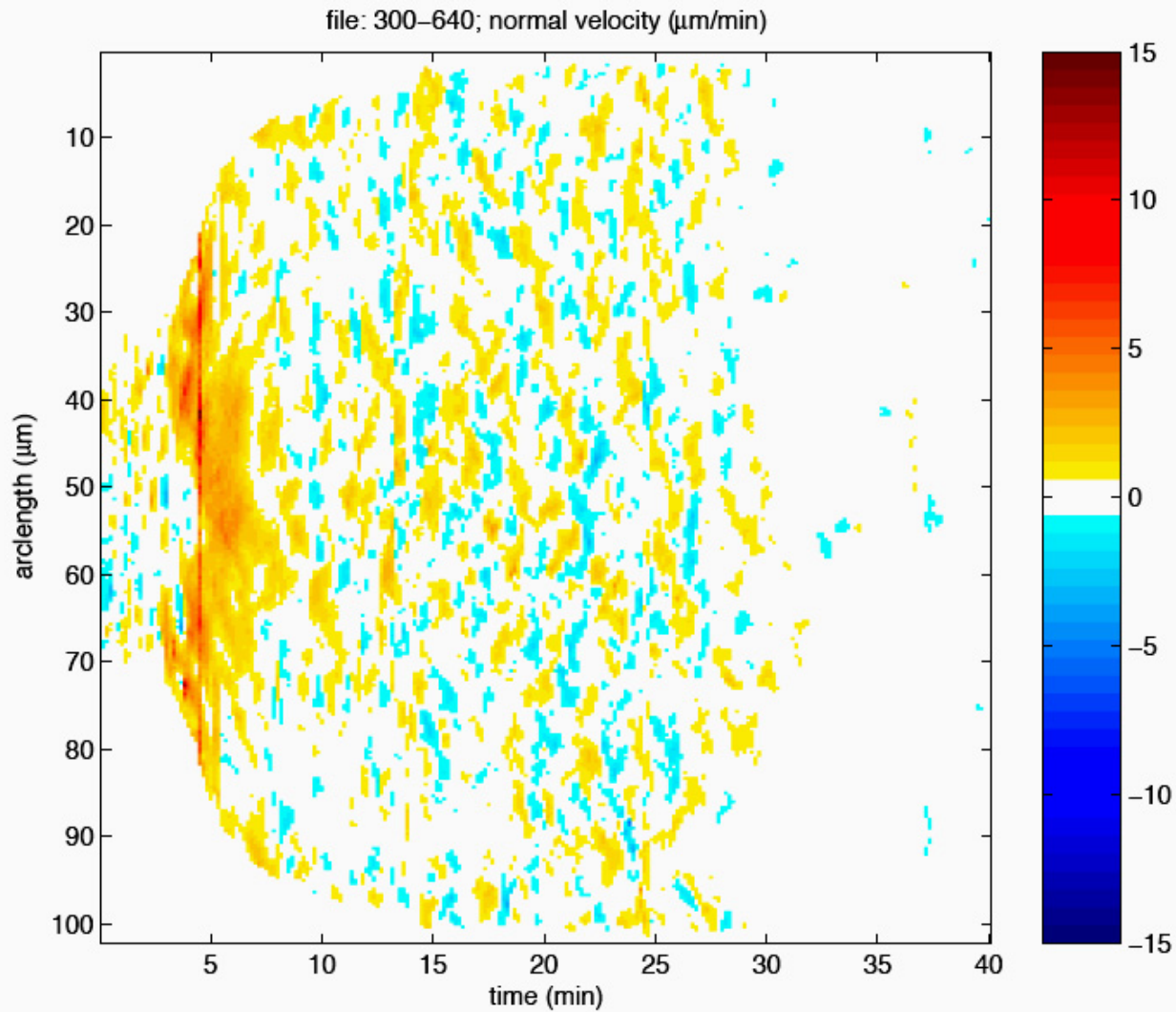
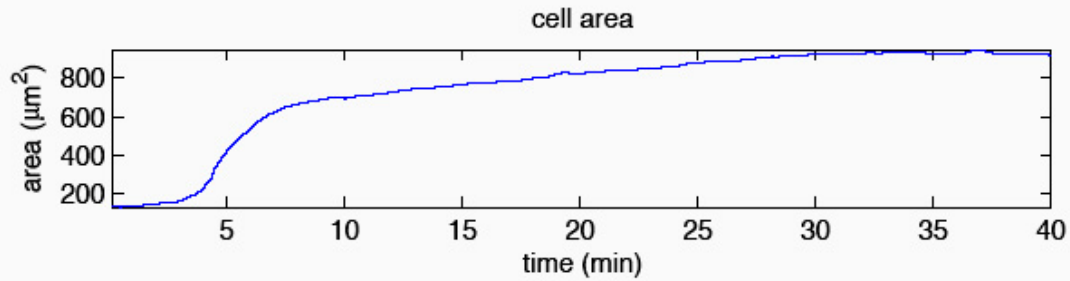


**C Anisotropic**

**D Isotropic**



MARCKS +/+

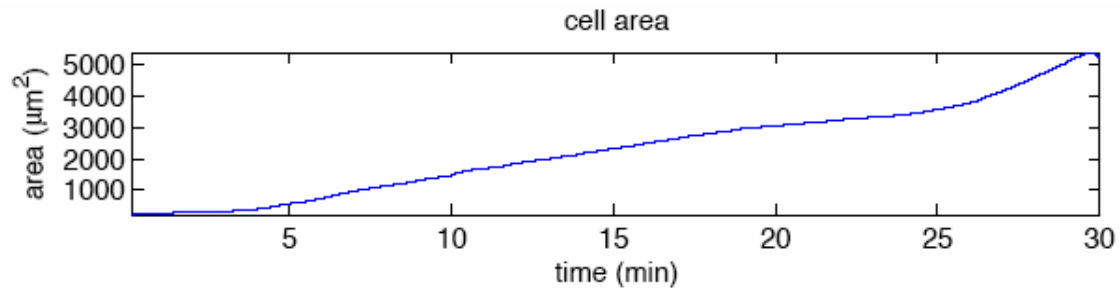


Harry Xenius

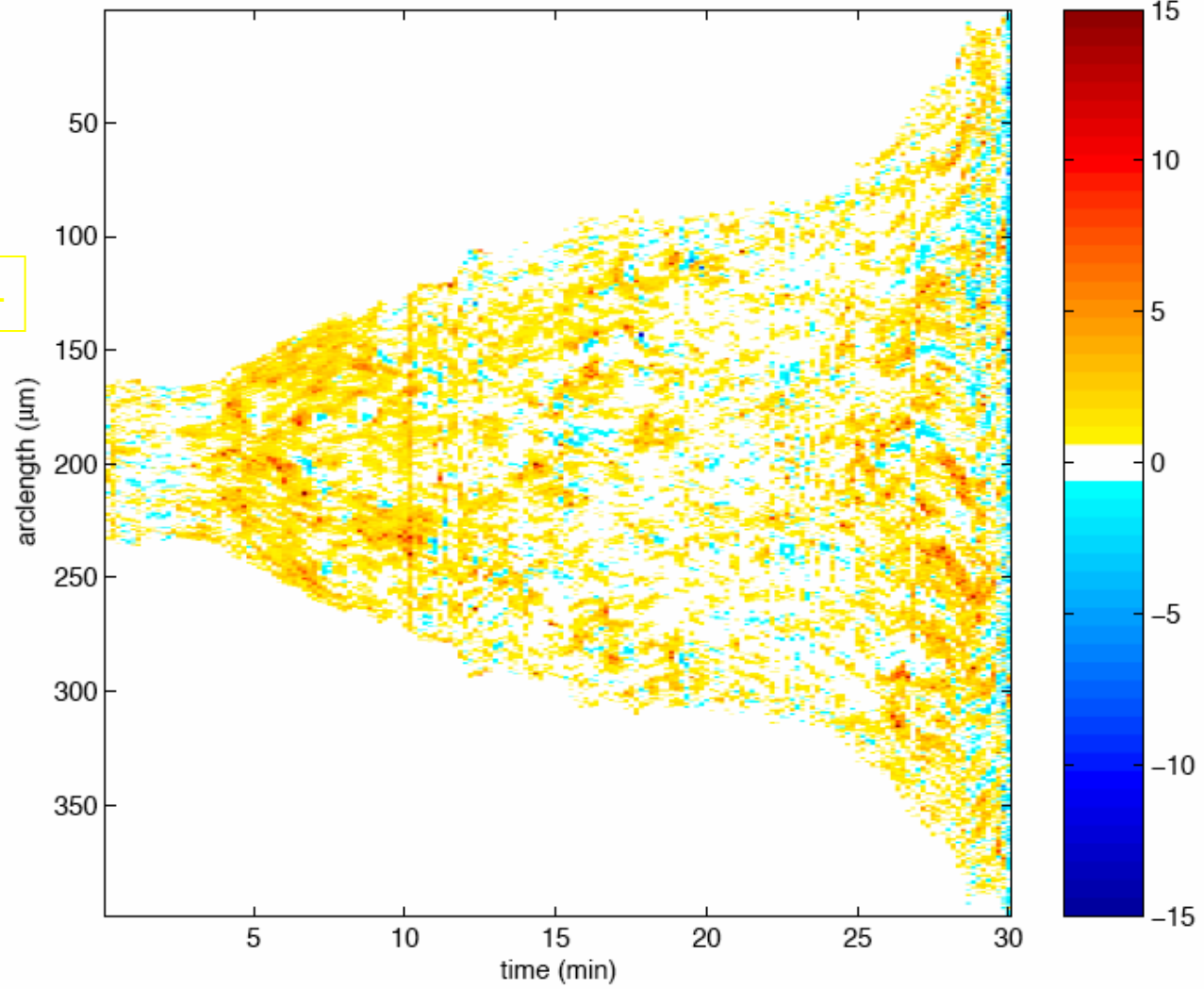
Collaborators

Chris Wiggins

Jake Hofman

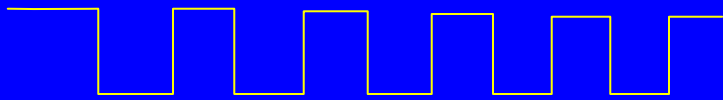


file: 340-210; normal velocity ( $\mu\text{m}/\text{min}$ )



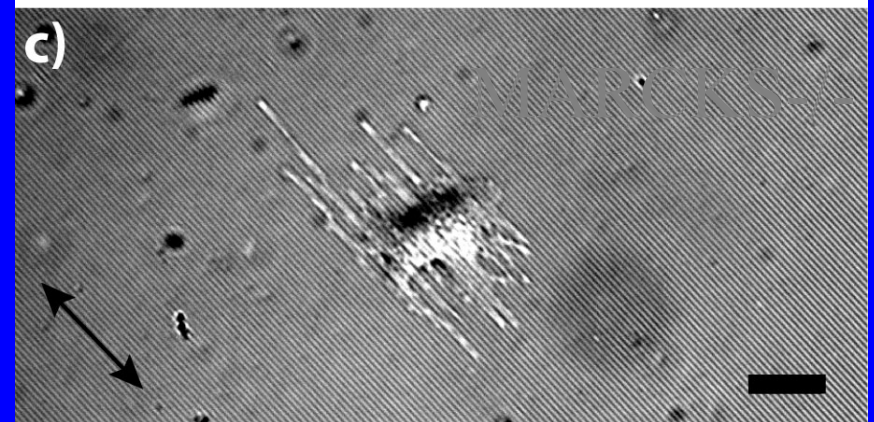
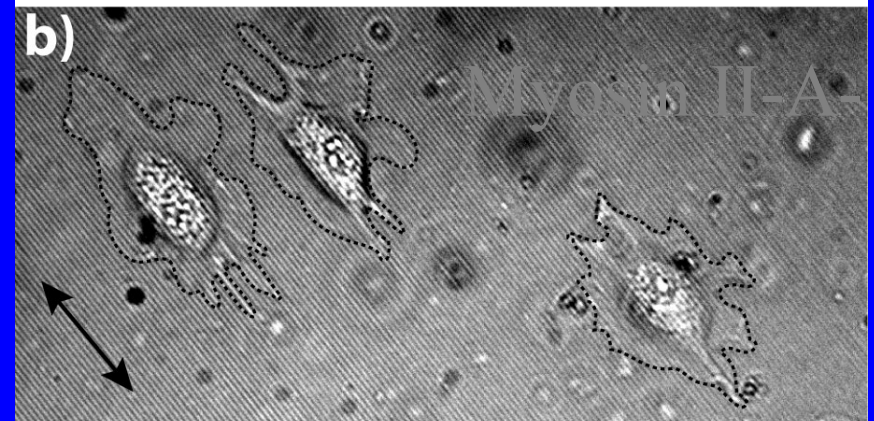
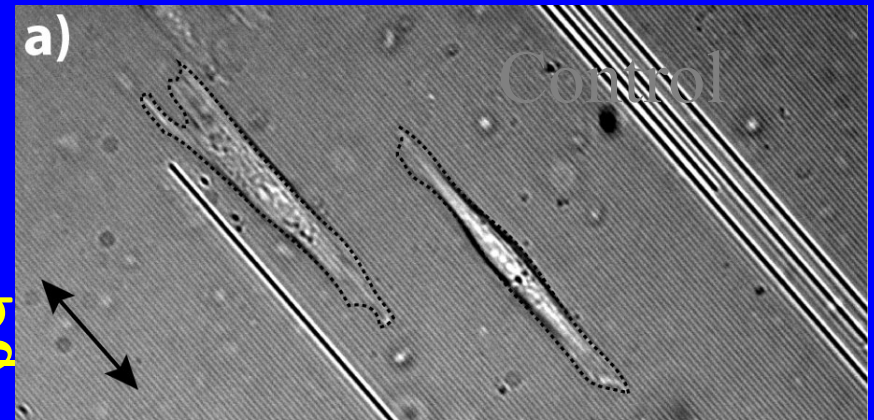
MARCKS-/-

# Regulation of Morphology by Substrate Engineering

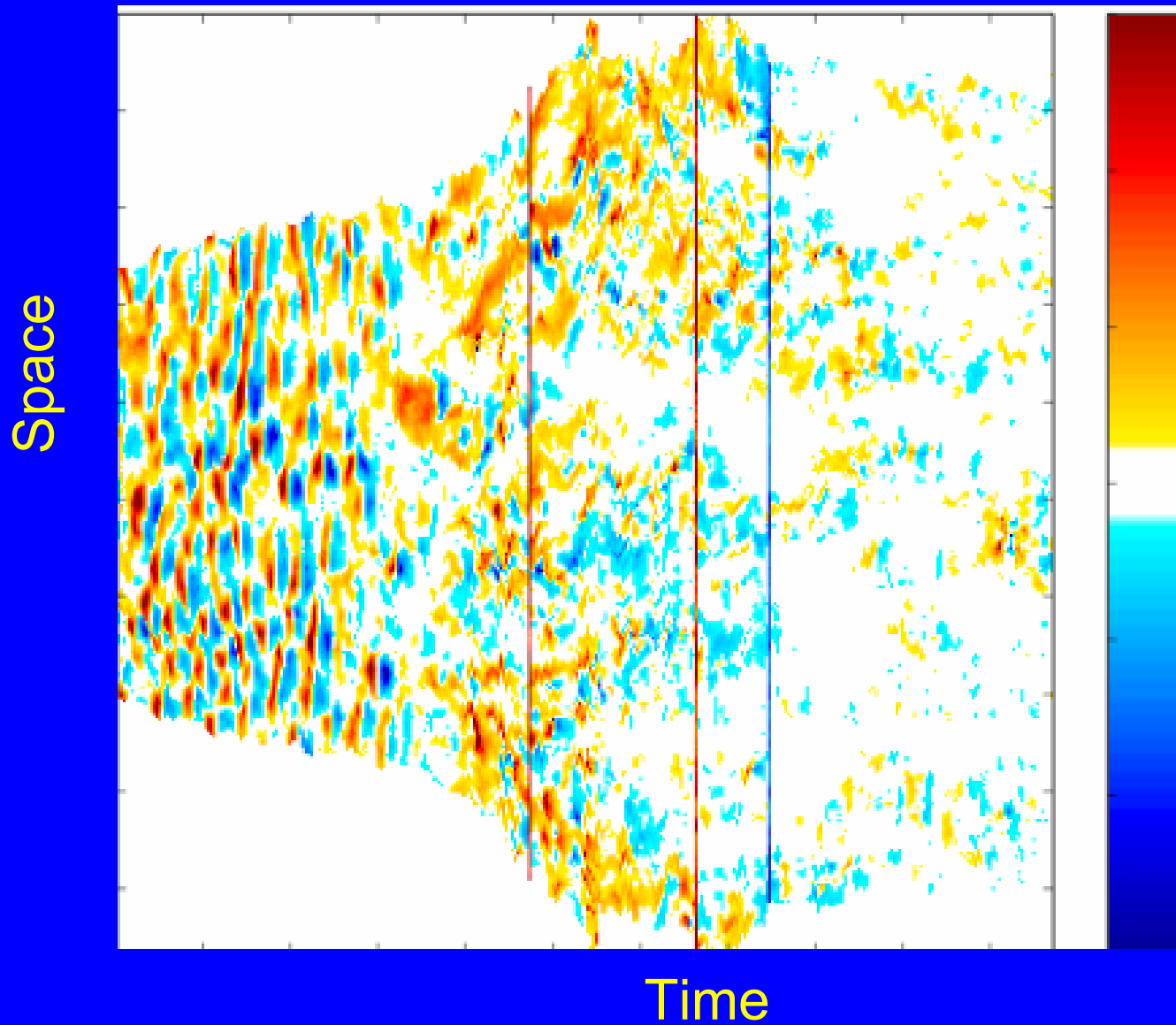


Substrates have  $1\mu\text{m}$  ridges separated by  $1\mu\text{m}$

Understanding the biochemical and biophysical basis of cell morphology



# Evidence for Spatial and Temporal Order



# Cross-Correlation Analyses in Space and Time

$v(t, s) \equiv$  membrane velocity matrix - average velocity

$T \equiv$  total time of  $v(t, s)$        $S \equiv$  total arclength of  $v(t, s)$

$\Delta t \equiv$  lag in time

$\Delta s \equiv$  lag in space

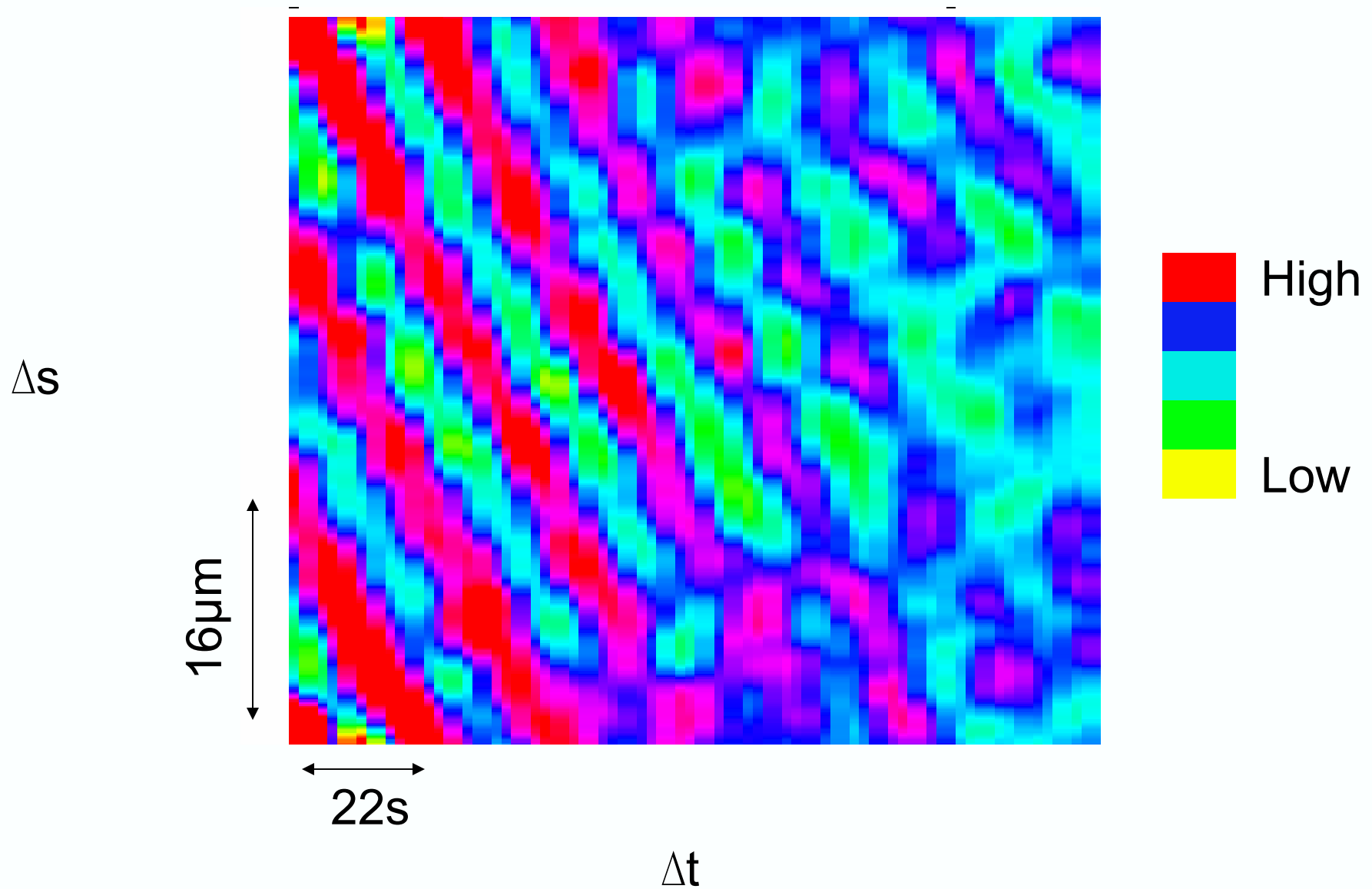
Correlation for a given point:

$$\text{correlation}(\Delta t, \Delta s) = v(i, j) * v(i + \Delta t, j + \Delta s)$$

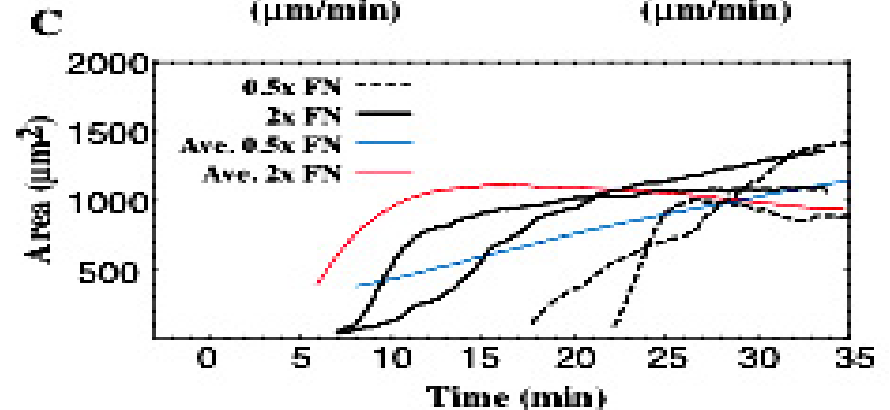
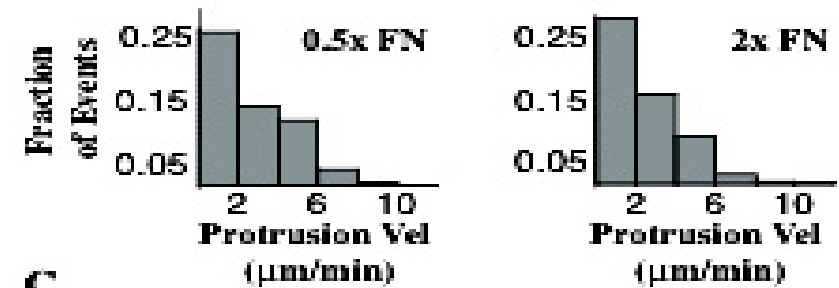
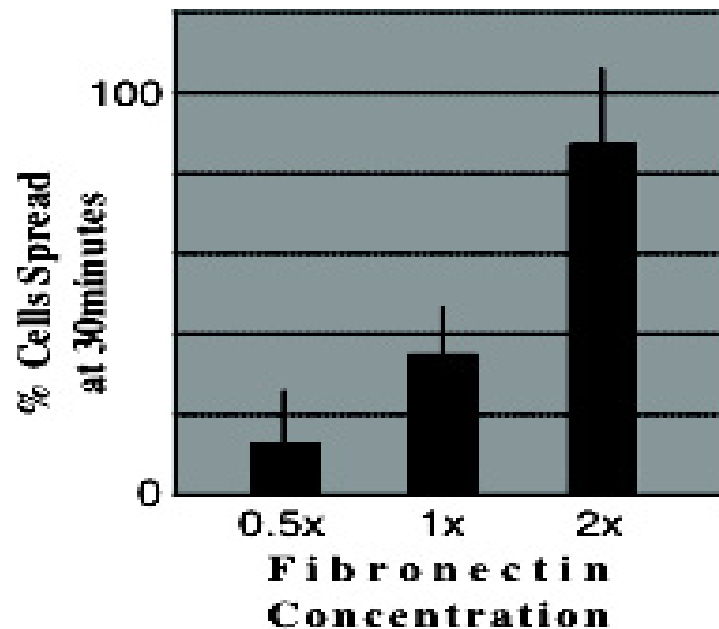
Correlation averaged over all points:

$$\text{correlation}(\Delta t, \Delta s) = \frac{\sum_{i=1}^{T-\Delta t} \sum_{j=1}^S v(i, j) * v(i + \Delta t, \text{mod}(j + \Delta s, S))}{(T - \Delta t) * S}$$

# Cross-Correlation Reveals Repeats at 16 $\mu\text{m}$ and 22s



# FN Concentration Affects Initiation, Not Rate of Spreading

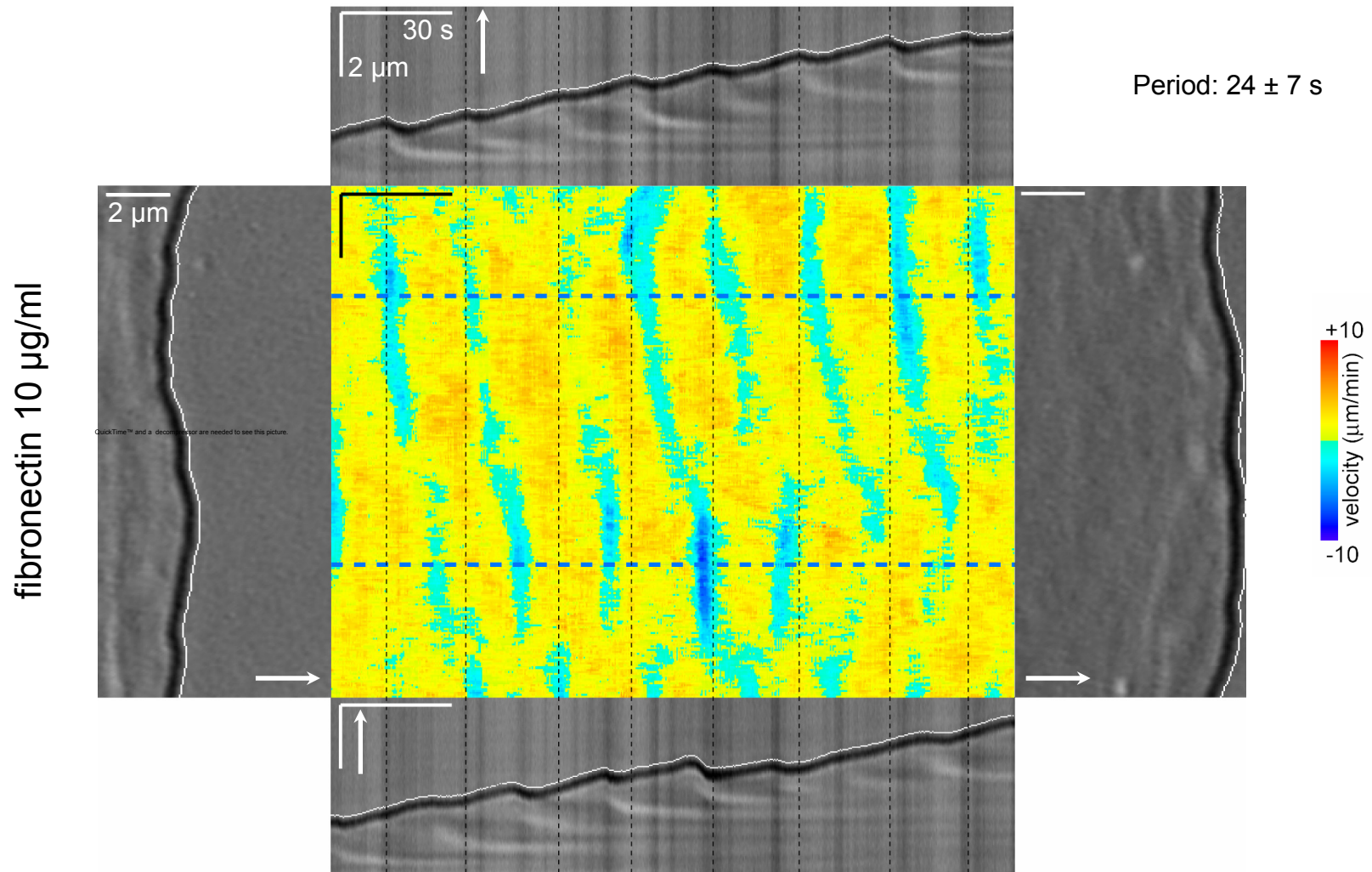




# Isotropic Spreading with GFP- $\alpha$ -actinin

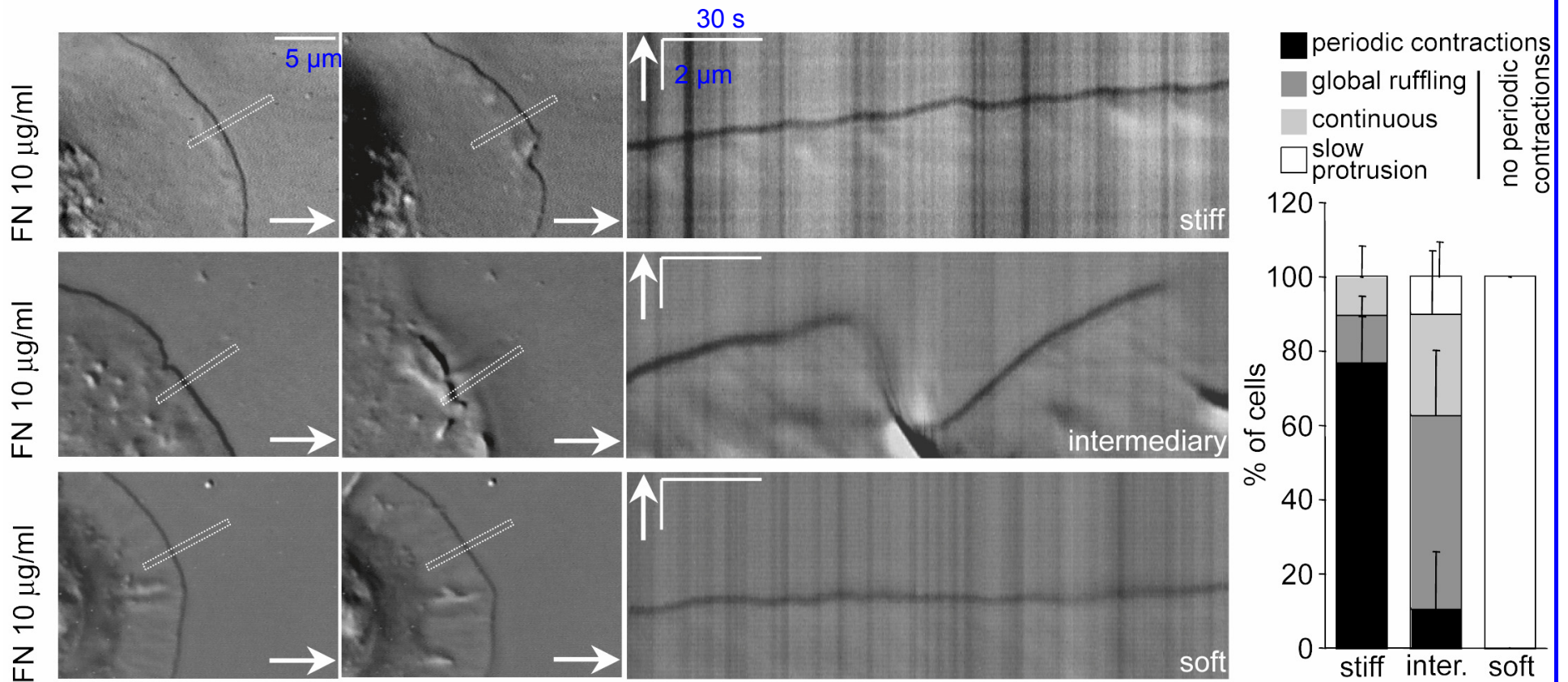
QuickTime™ and a Video decompressor are needed to see this picture.

# Local periodic retractions of the cell edge during lamellipodial extension

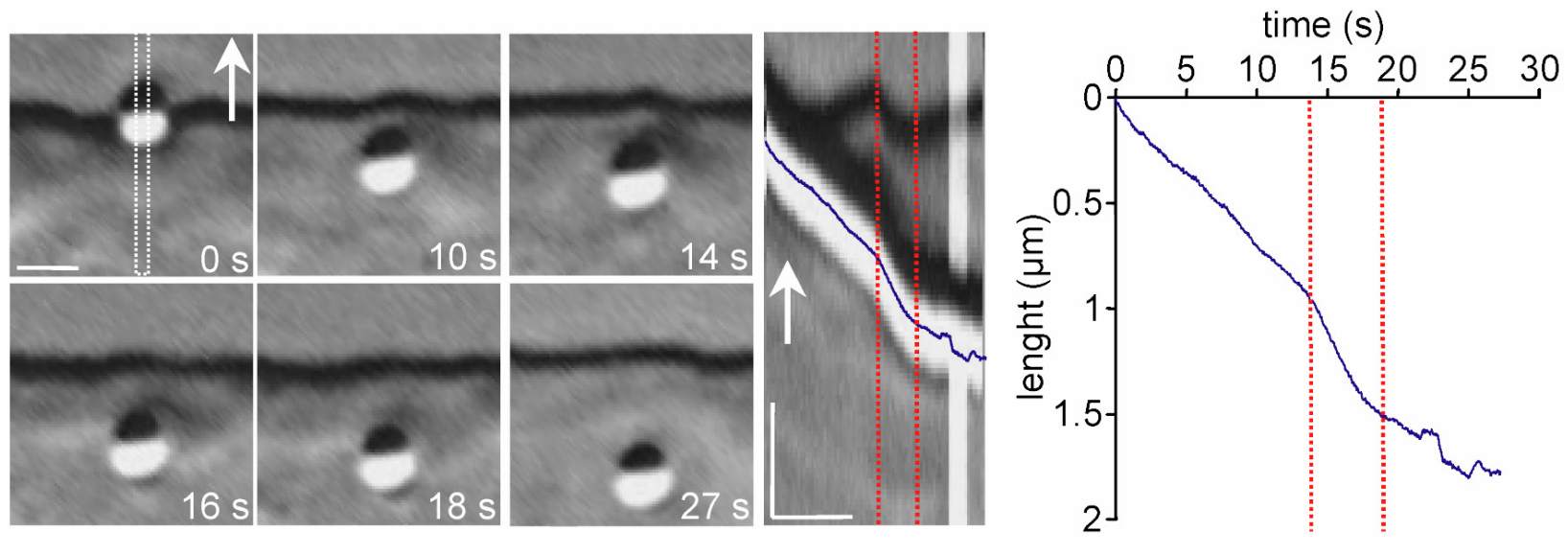


Giannone et al., (2004) Cell. 116:431-43.

# Periodic contractions depend on a stiff substrate

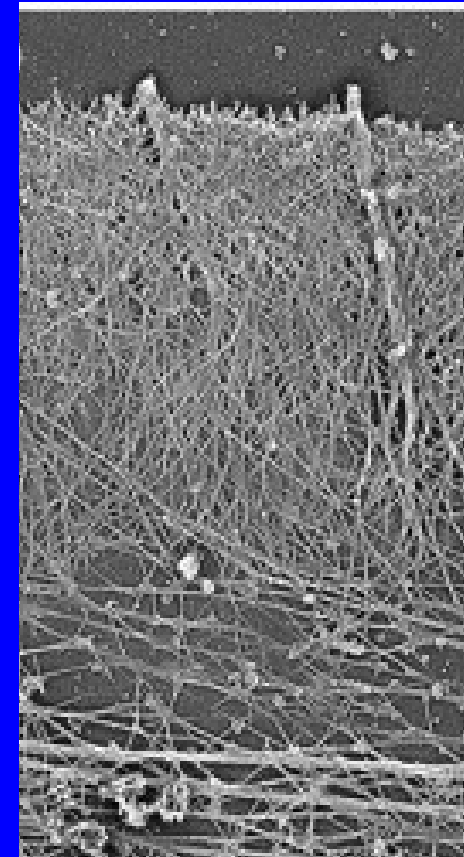
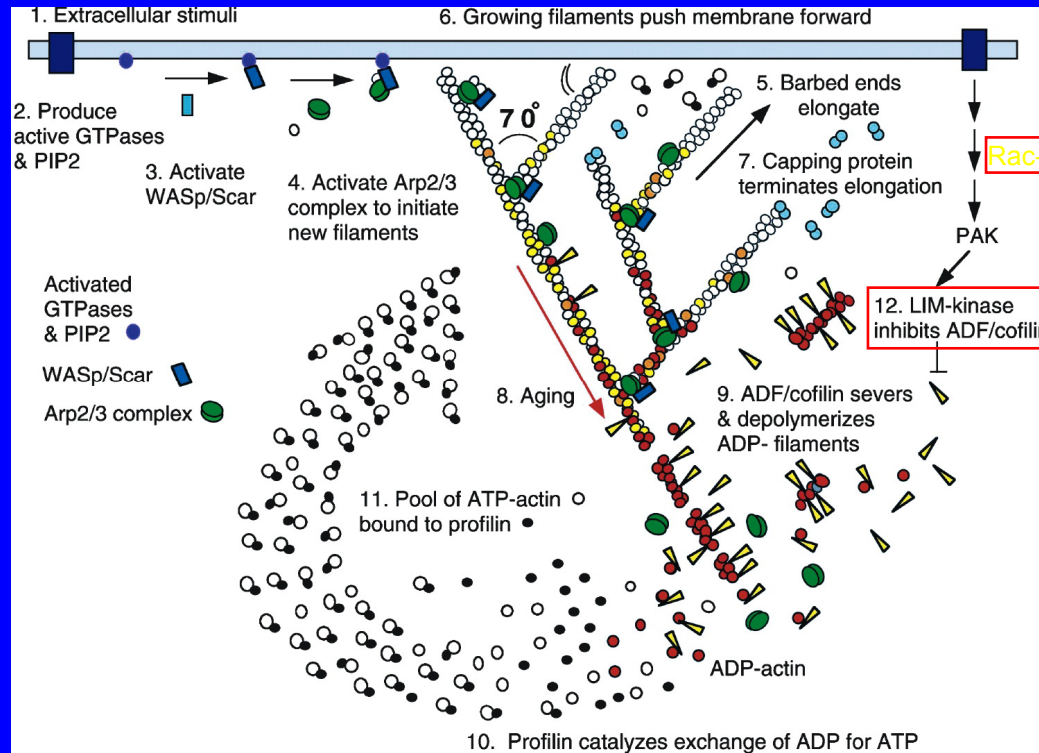


# Periodic Retractions Are Periodic Contractions of the Lamellipodia



QuickTime™ and a decompressor are needed to see this picture.

# Lamellipodial extension driven by actin assembly



Lamellipodia

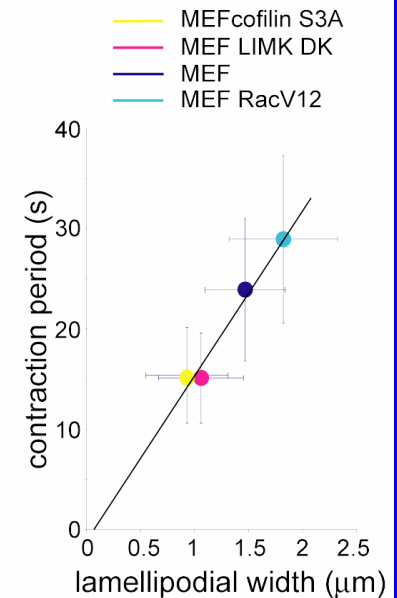
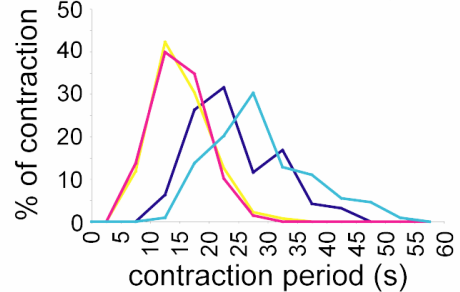
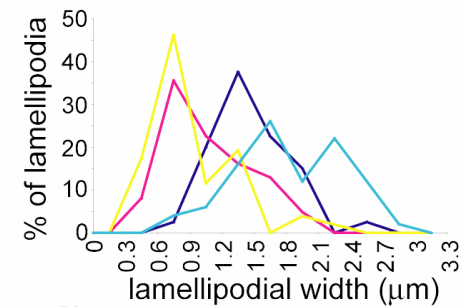
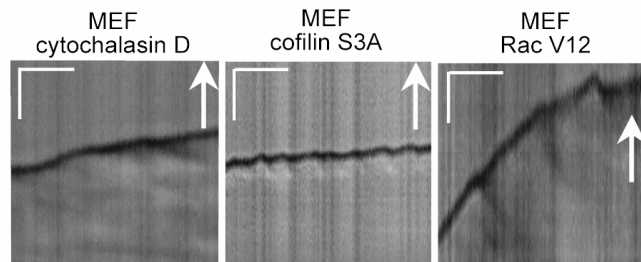
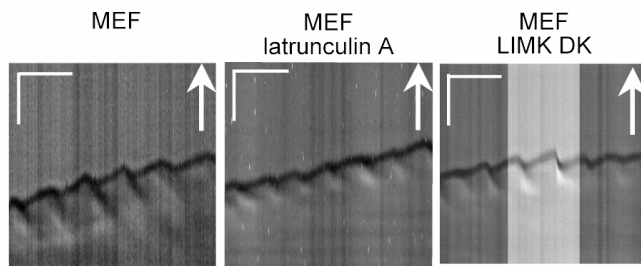
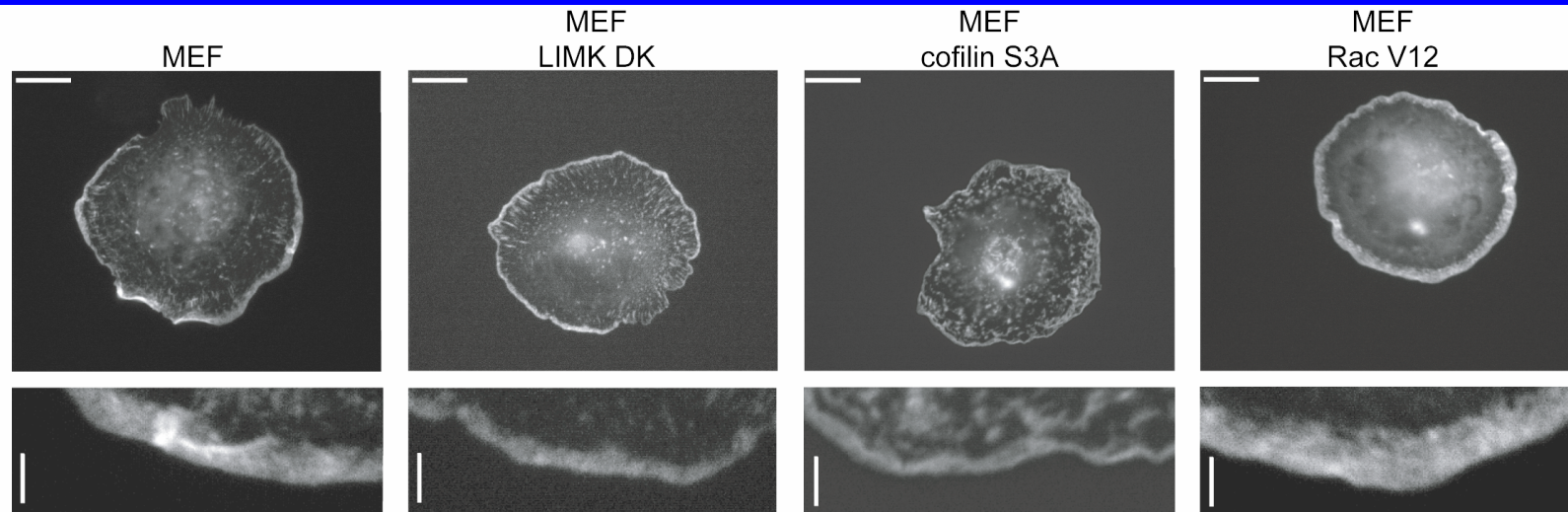
Lamella

Forces generated by Myosins?

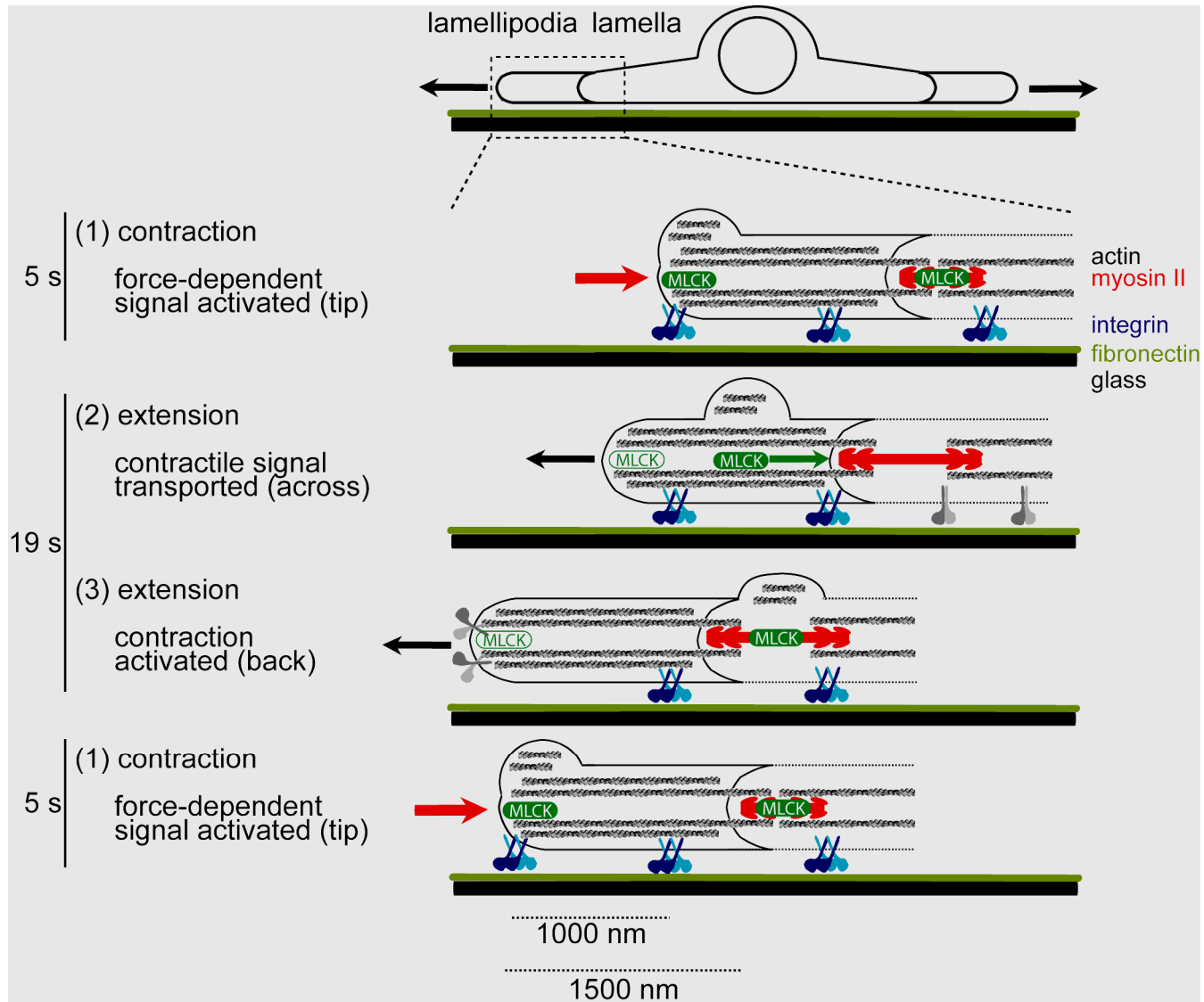
Actin polymerization (front) and depolymerization (rear) define the lamellipodial width

Cell spreading and migration require myosin activity

# Period Set by Width of Actin Meshwork



# Model of signaling by cytoskeletal transport



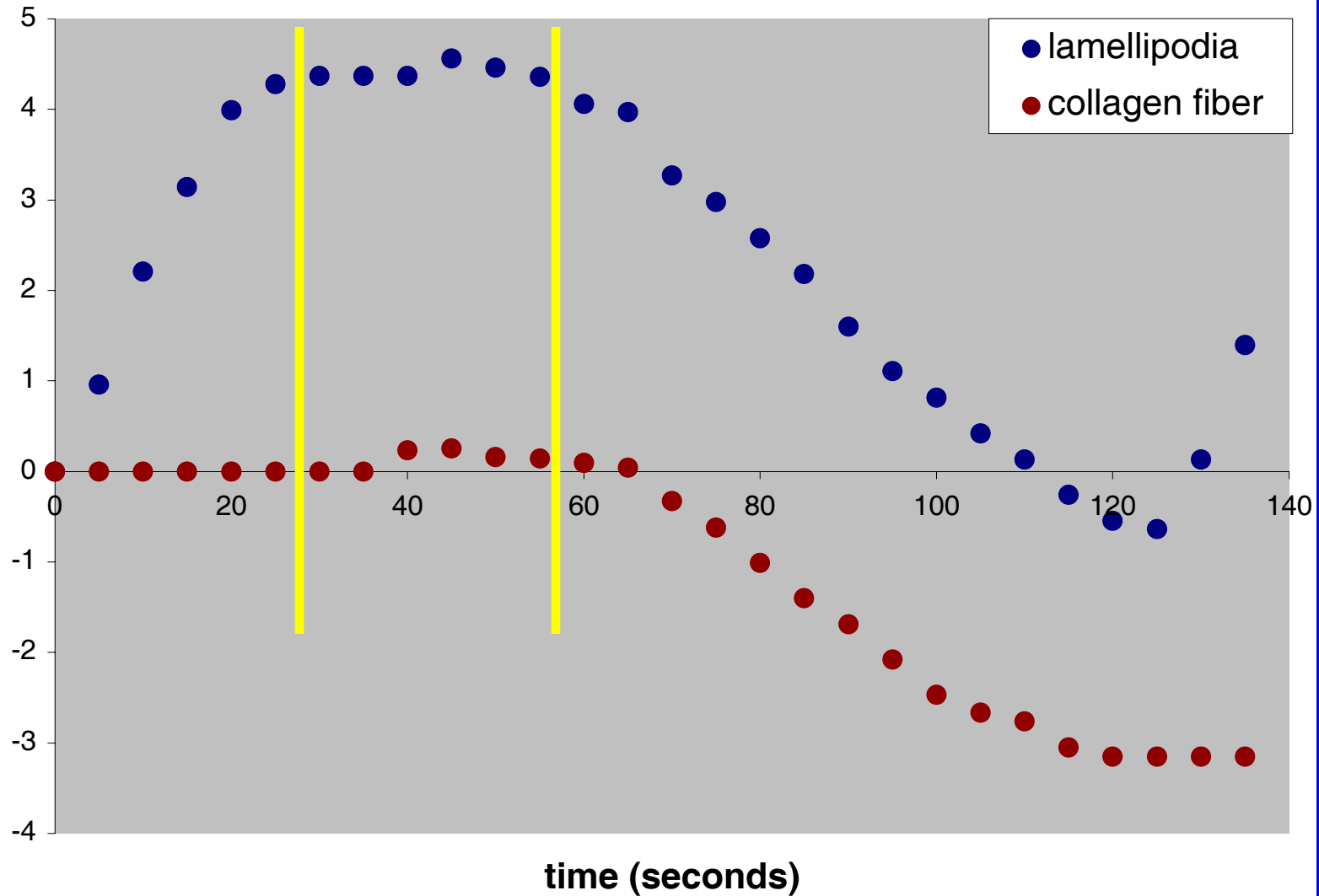
# Hand-Over-Hand Movement of Collagen Fibers by Myosin II-B

QuickTime™ and a Video decompressor are needed to see this picture.

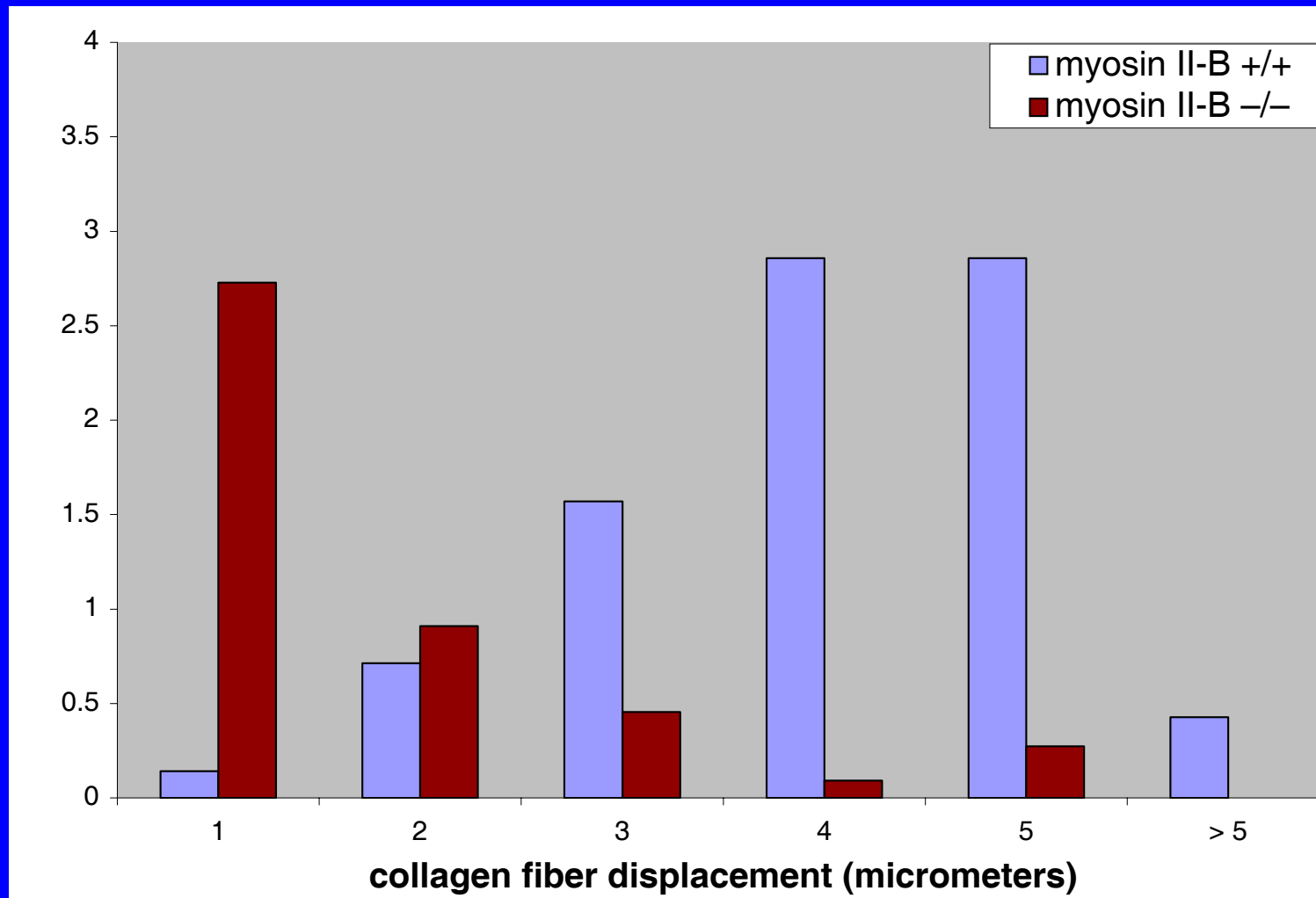
Meshel, Wei, Adelstein & Sheetz, 2005 Nat. Cell Biol. 7:157



# 1 cycle of motility = “event”



# II-B $-/-$ cells have slower, shorter events

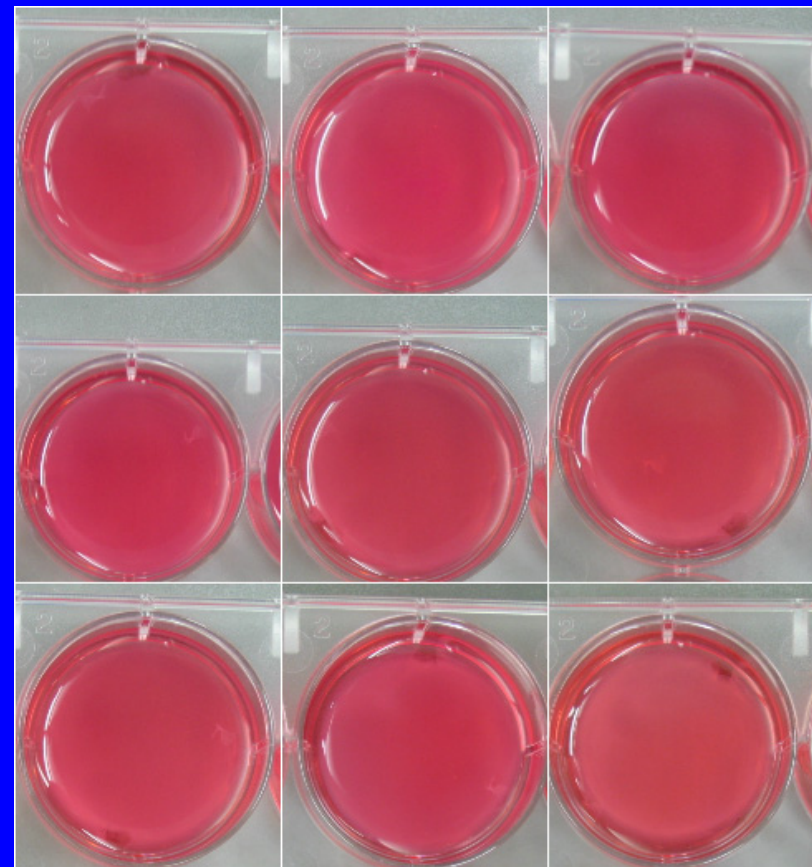
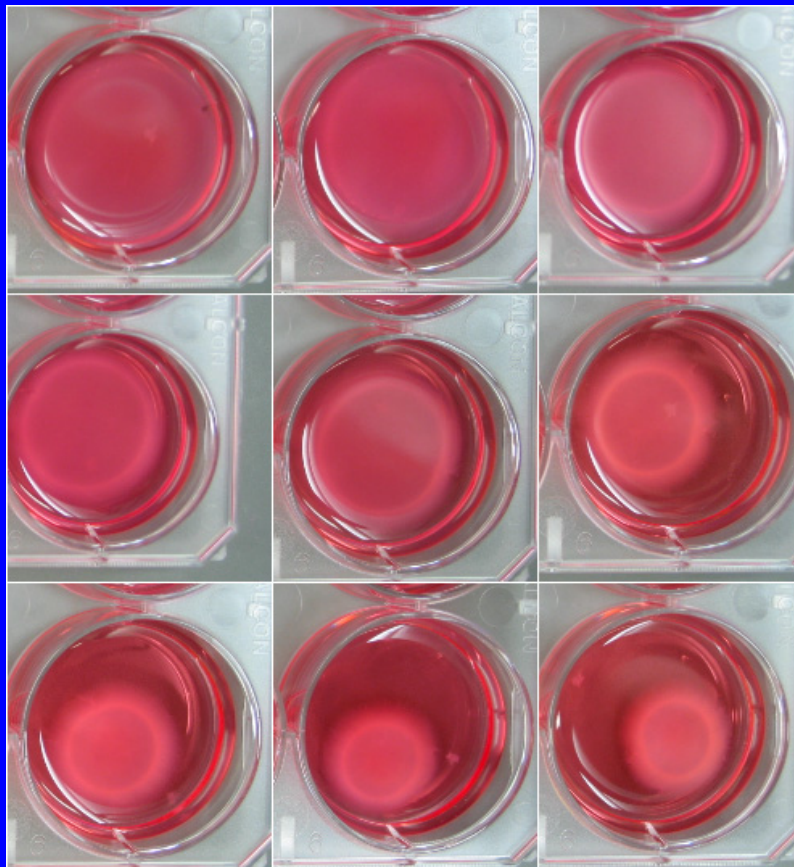


\*retransfection of GFP-NMHC II-B restores function while GFP-II-A does not...

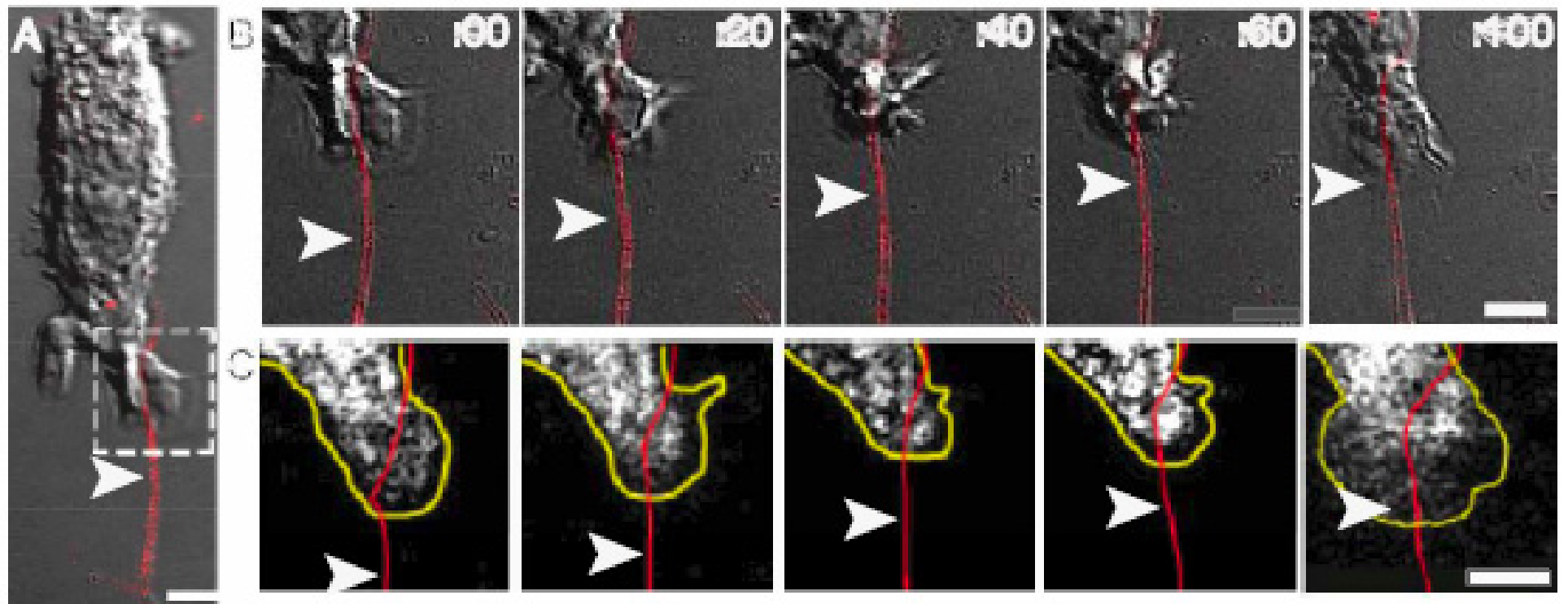
# Myosin II-B is required for 3-D collagen gel contraction

wild type

myosin II-B -/-

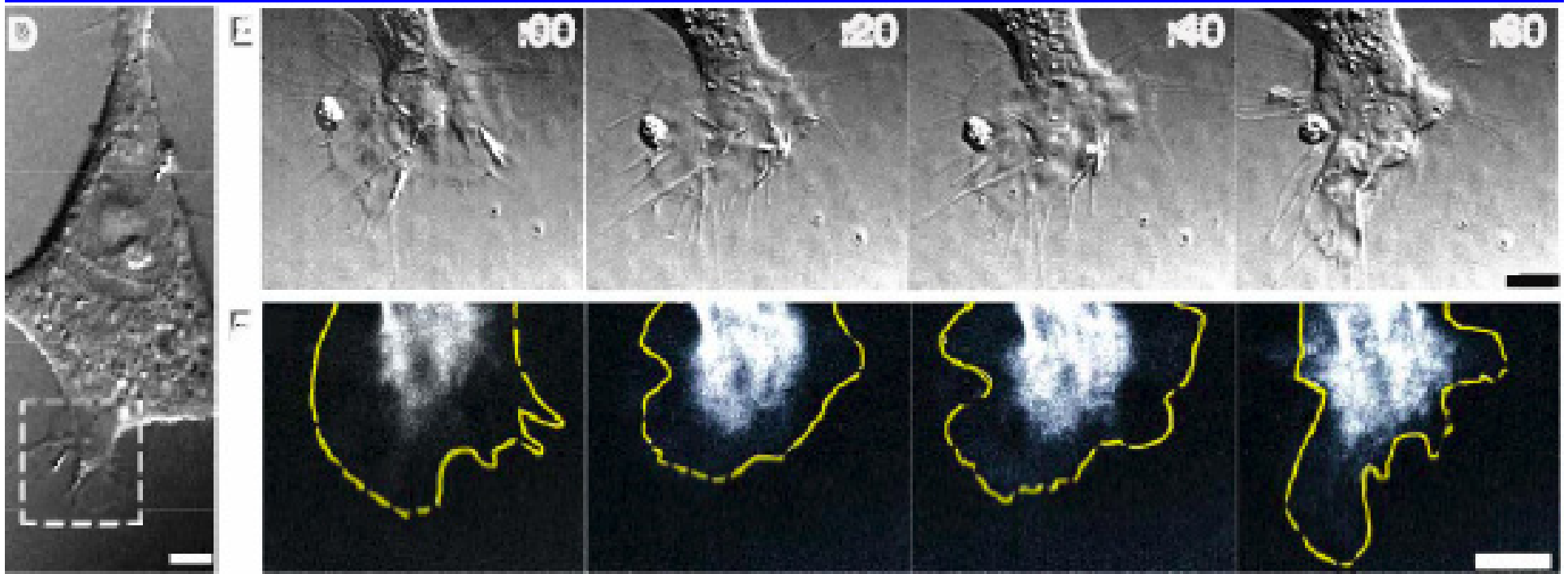


# Myosin II-B Assembles in Lamellipodia & Moves With Fiber



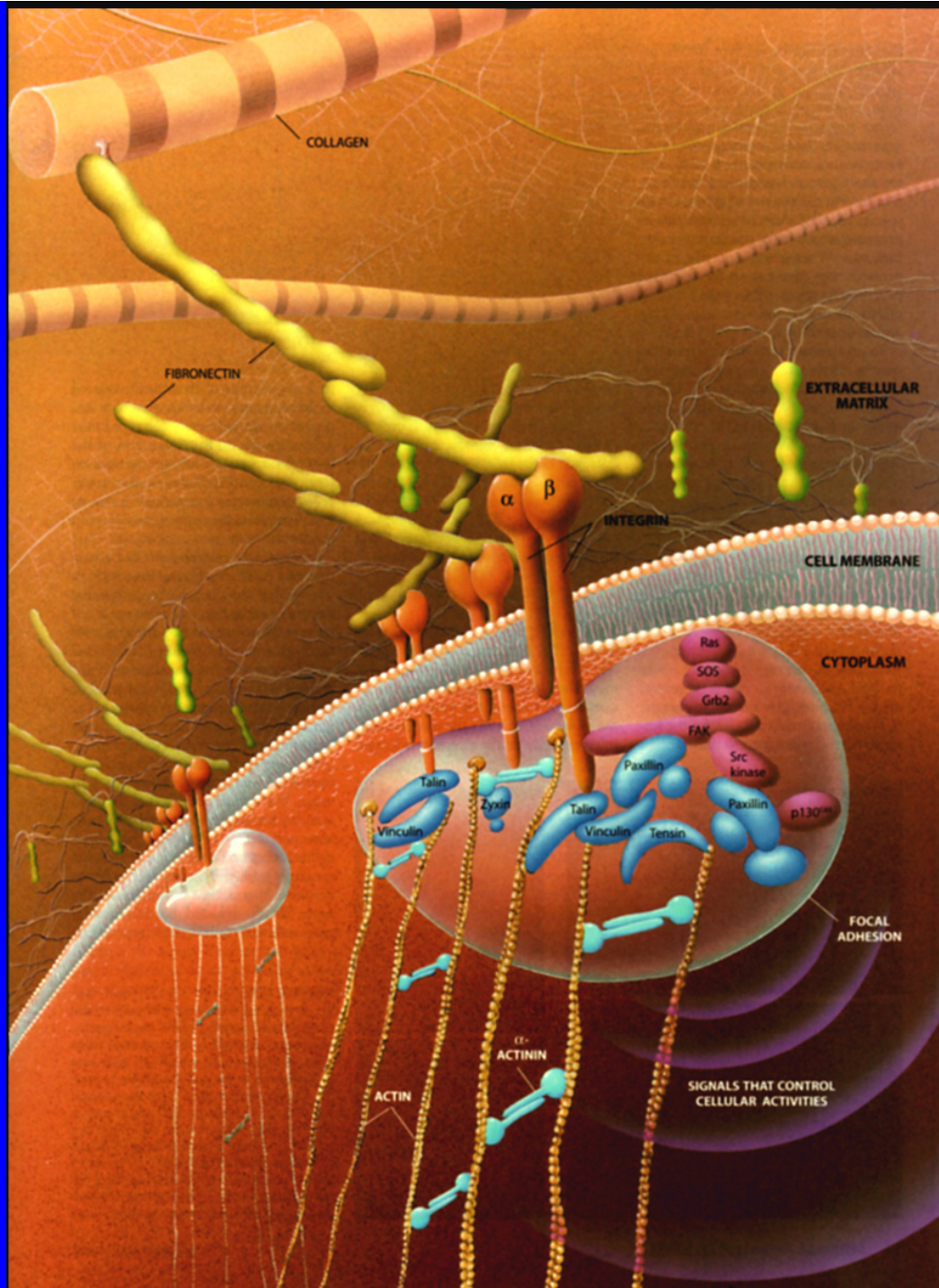
Meshel, Wei, Adelstein & Sheetz, 2005 Nat. Cell Biol. 7:157

# Myosin II-B Is Not in Lamellipodia on 2-D Collagen



# General Lessons for Robust Functions at Submicron Level

- Compartmentalization and Modular design
- Spatial Ordering of Multiple sites for Specificity
- Functional Activity Turns Off Automatically
- Multiple Activity Cycles for Important Functions
- Many Low Fidelity Steps for High Fidelity Process



Horwitz, Scientific American

# Acknowledgements

## Sheetz Lab

Drazen Raucher

Catherine Galbraith

Dan Felsenfeld

Daniel Choquet

Ben Dubin-Thaler

Yasuharo Sawada

Jianwu Dai

Goetz Wichert

Julia Sable

Guoying Jiang

Adam Meshel

Masako Tamada

## Duke University

Dr. Robert Hochmuth

Stuart McLaughlin  
(SUNY Stonybrook)

Michael Edidin

## North Carolina State University

Dr. Paul Franzon

David Nackashi